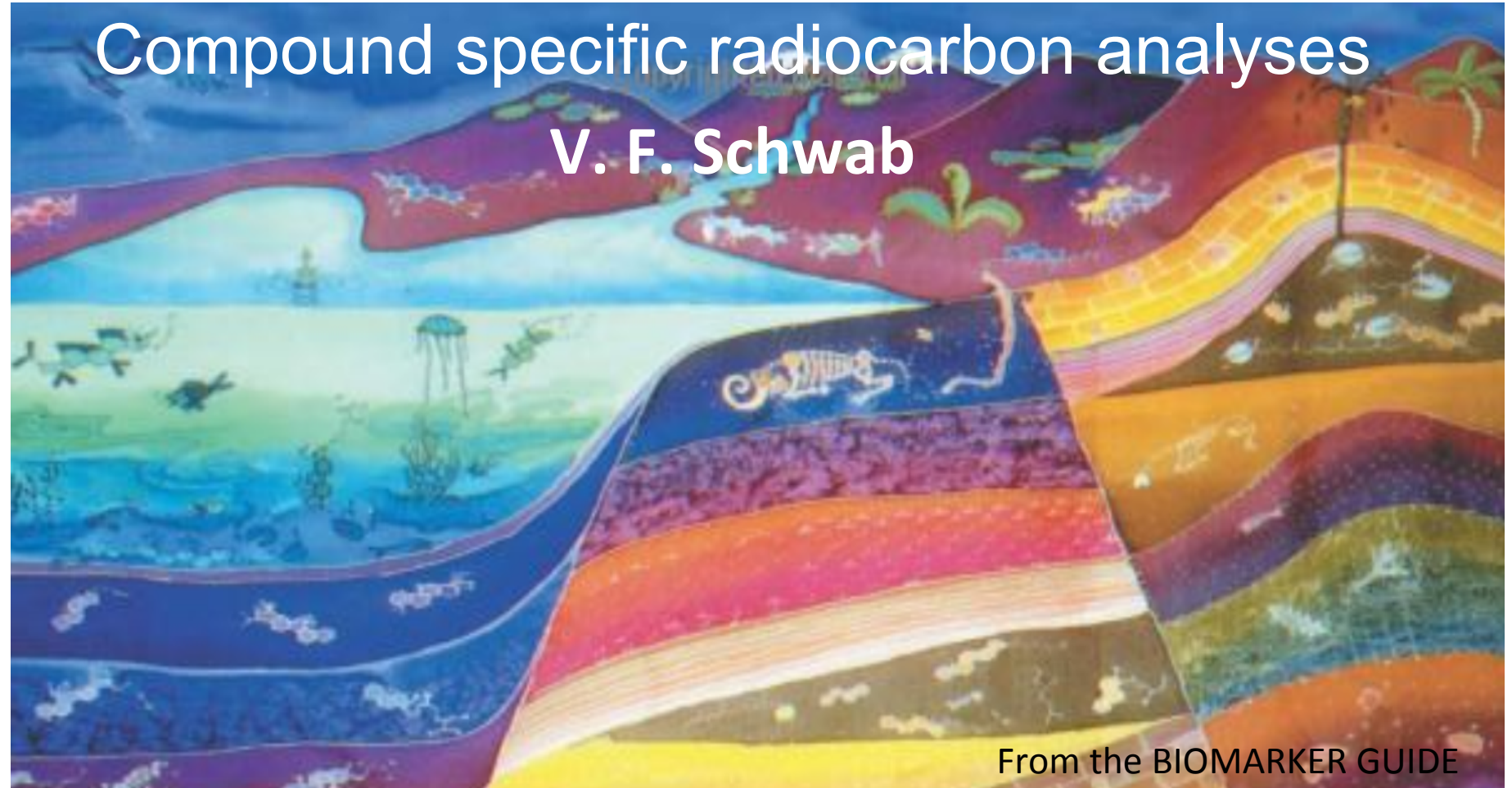


RADIOCARBON IN THE EARTH SYSTEM

Compound specific radiocarbon analyses

V. F. Schwab



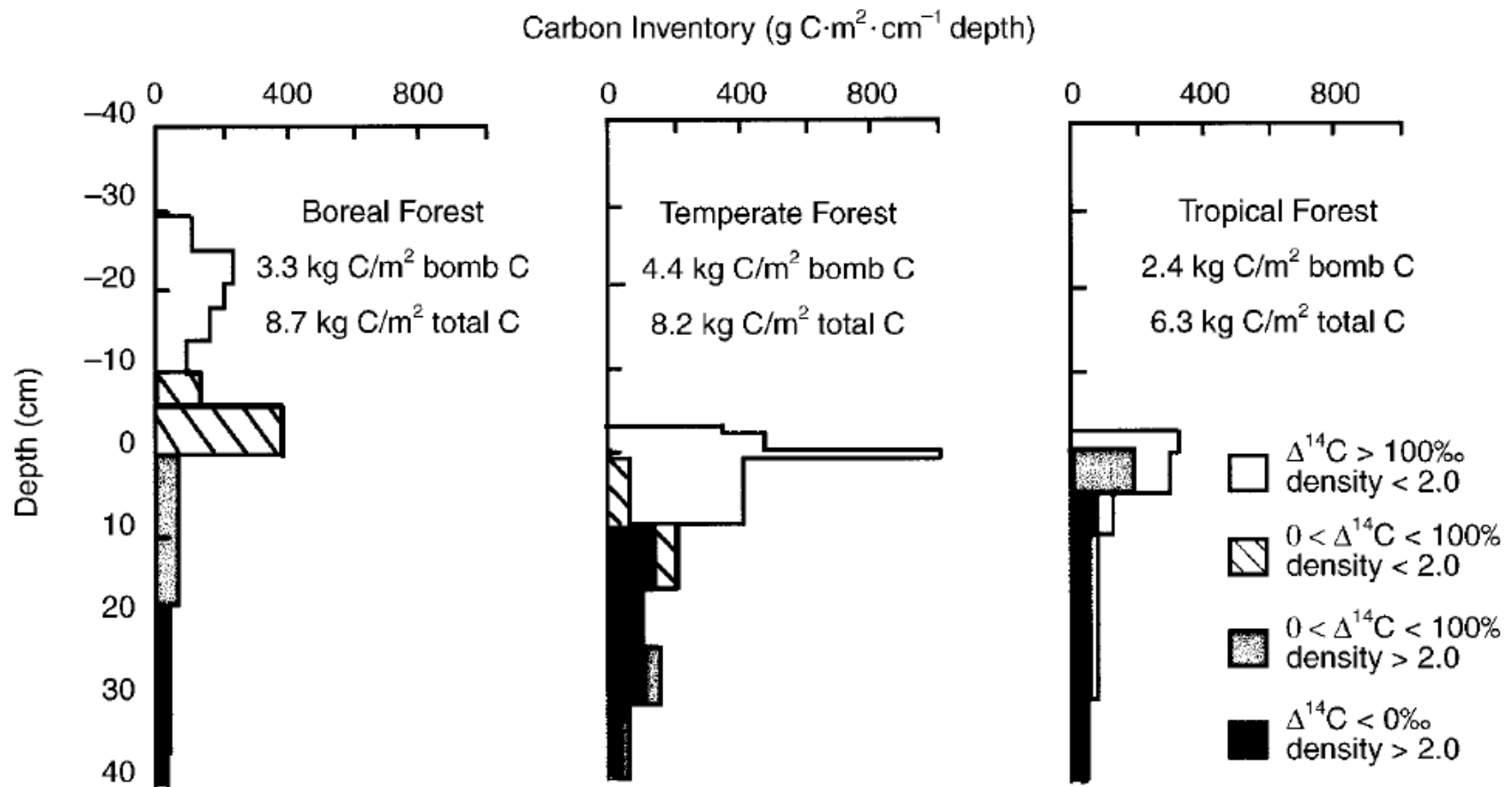
From the BIOMARKER GUIDE



seit 1558

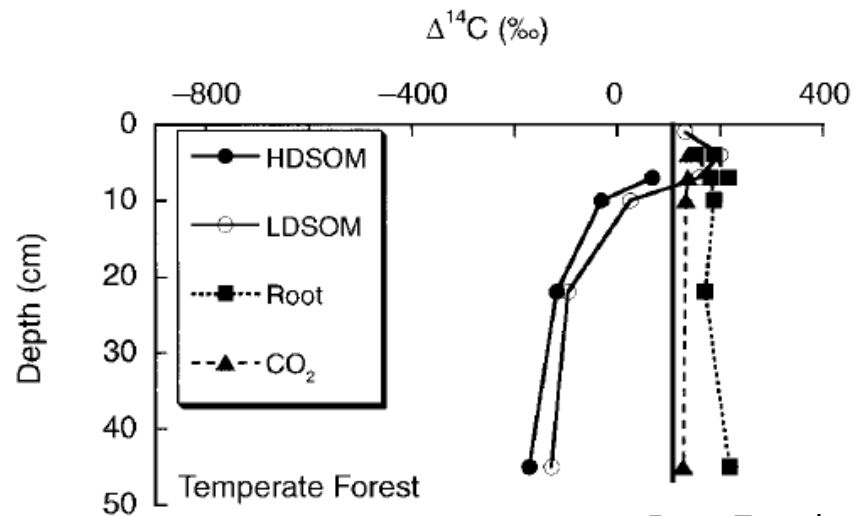
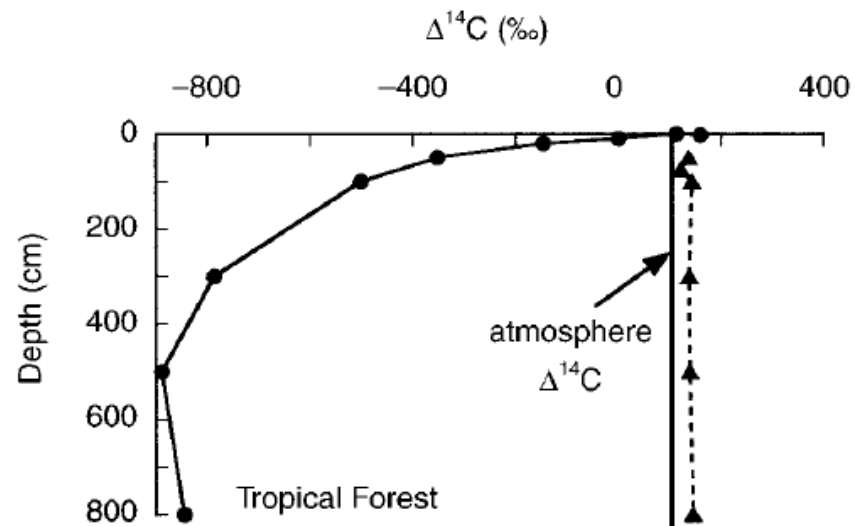


Example in age variations of organic carbon pools in soils



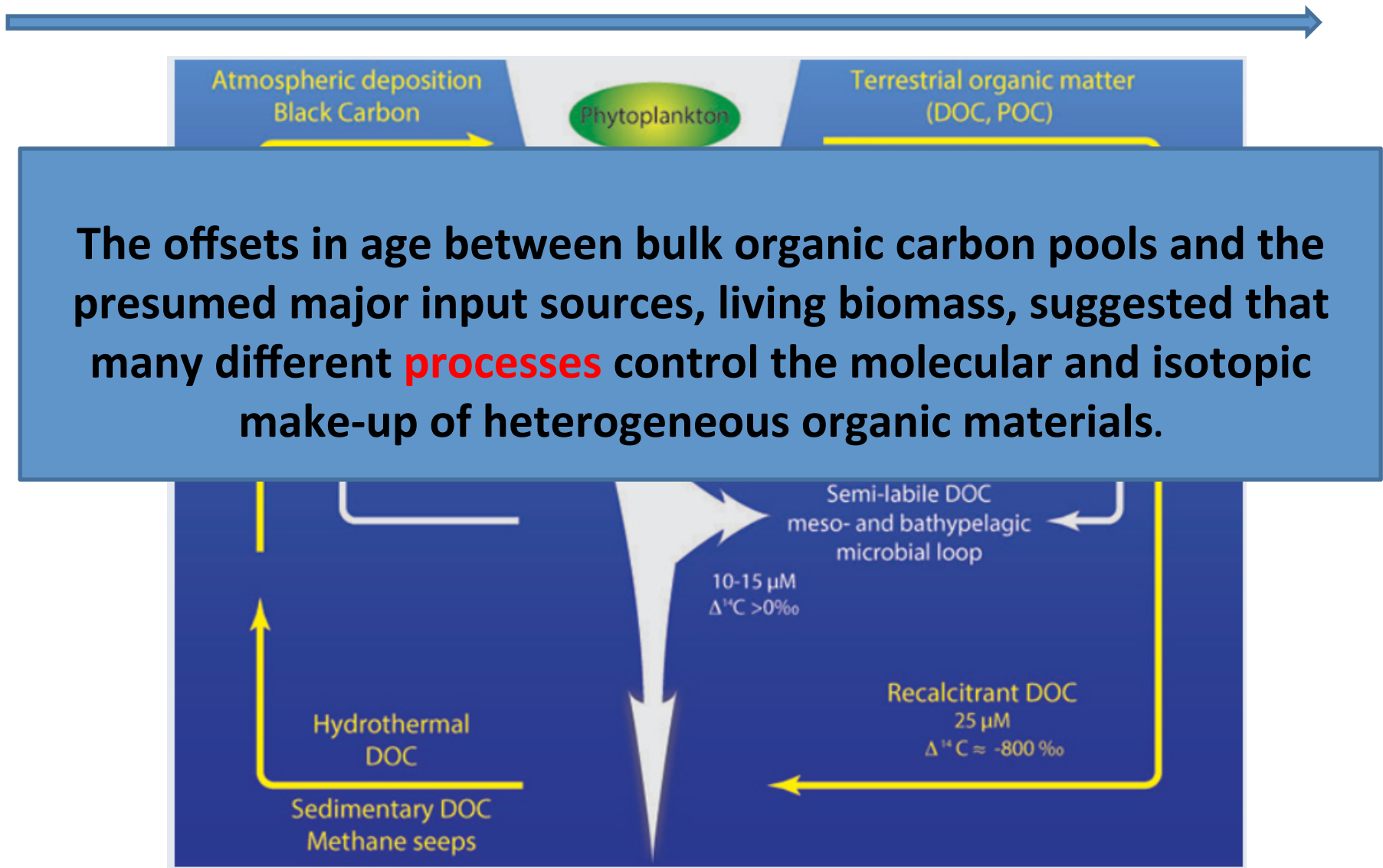
From Trumbore et al., 2000 (*Ecological Applications*)

Example in age variations of organic carbon pools in soils

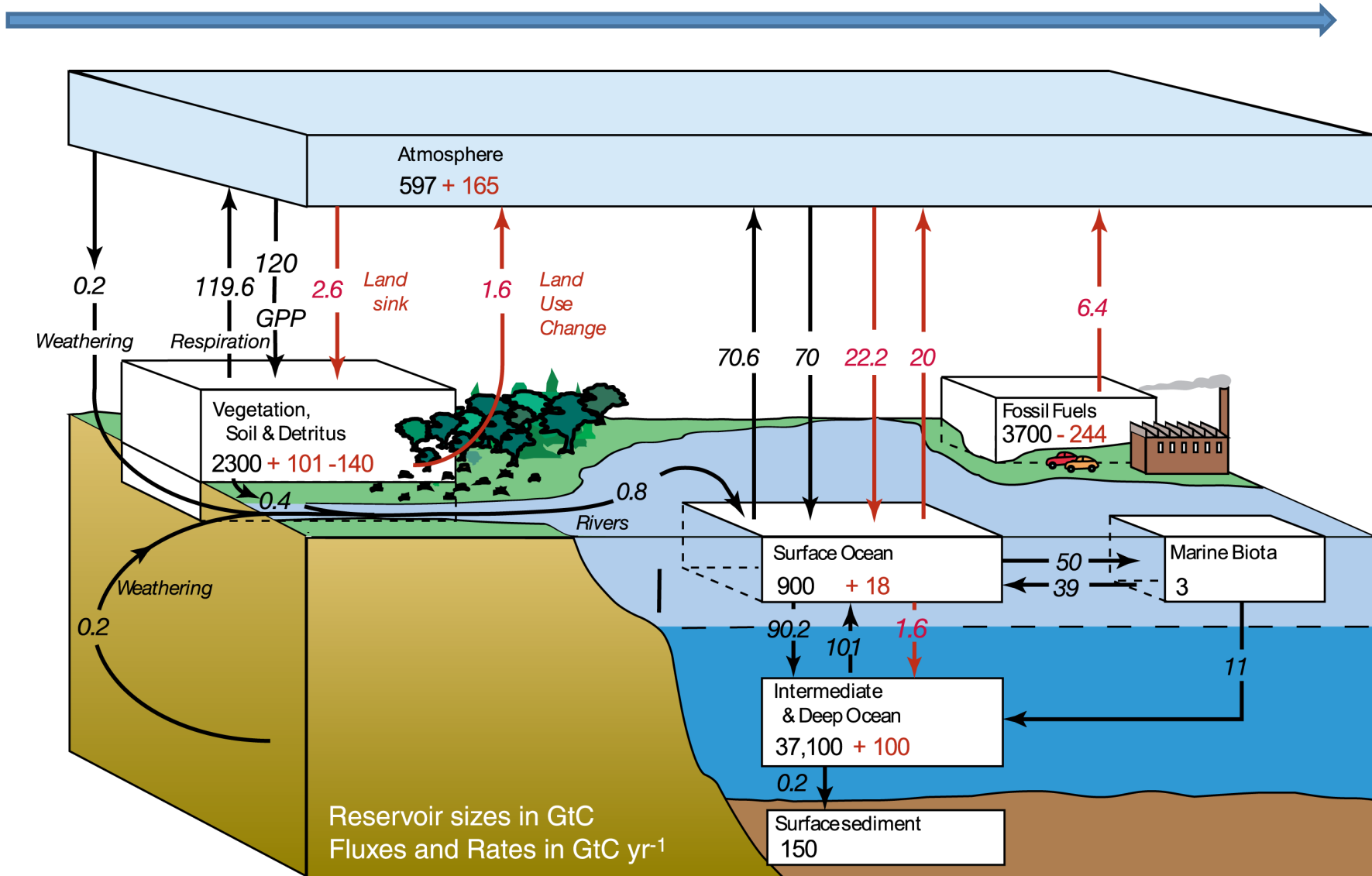


From Trumbore et al., 2000 (*Ecological Applications*)

Large heterogeneity in oceanic DOC and POC



The Anthropogenic Carbon Cycle



<https://www.gfdl.noaa.gov/anthropogenic-carbon-cycle/>

compound-specific radiocarbon analysis



Following on the heels of compound-specific stable isotope analysis (GC-C-IRMS) compound-specific radiocarbon analysis (CSRA) is envisioned as a way to couple the diversity of carbon sources with the residence time of carbon in the respective source pools. (Ingalls and Pearson.

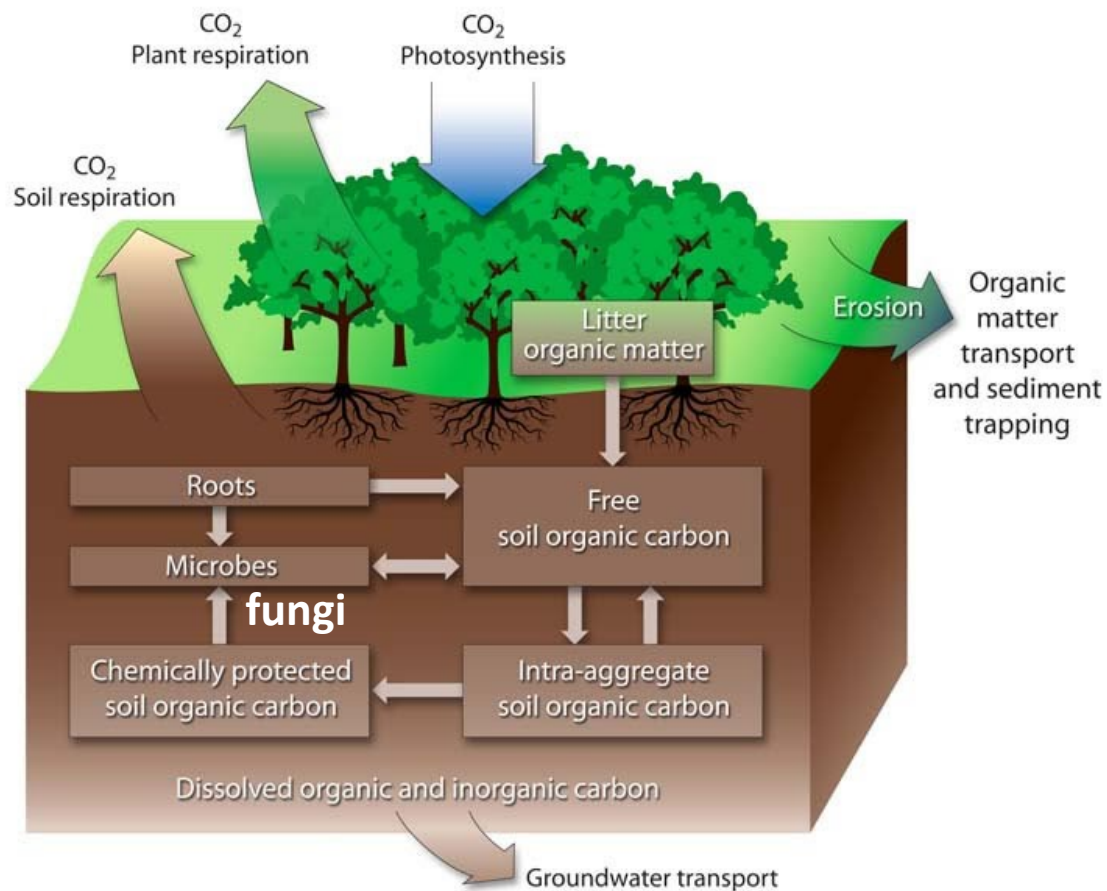
Oceanography (2005))

Applications of CSRA include studies of the



- **fixation, transformation, transport, and preservation of organic carbon**
- **elucidation of microbial metabolic pathways**
- **sources and reactivity of dissolved organic carbon**
- **organic paleo-proxy dating**
- **development of improved sediment chronologies.**

Single-compound radiocarbon analysis help to couple the diversity of carbon sources with the residence time of carbon in the respective source pools.



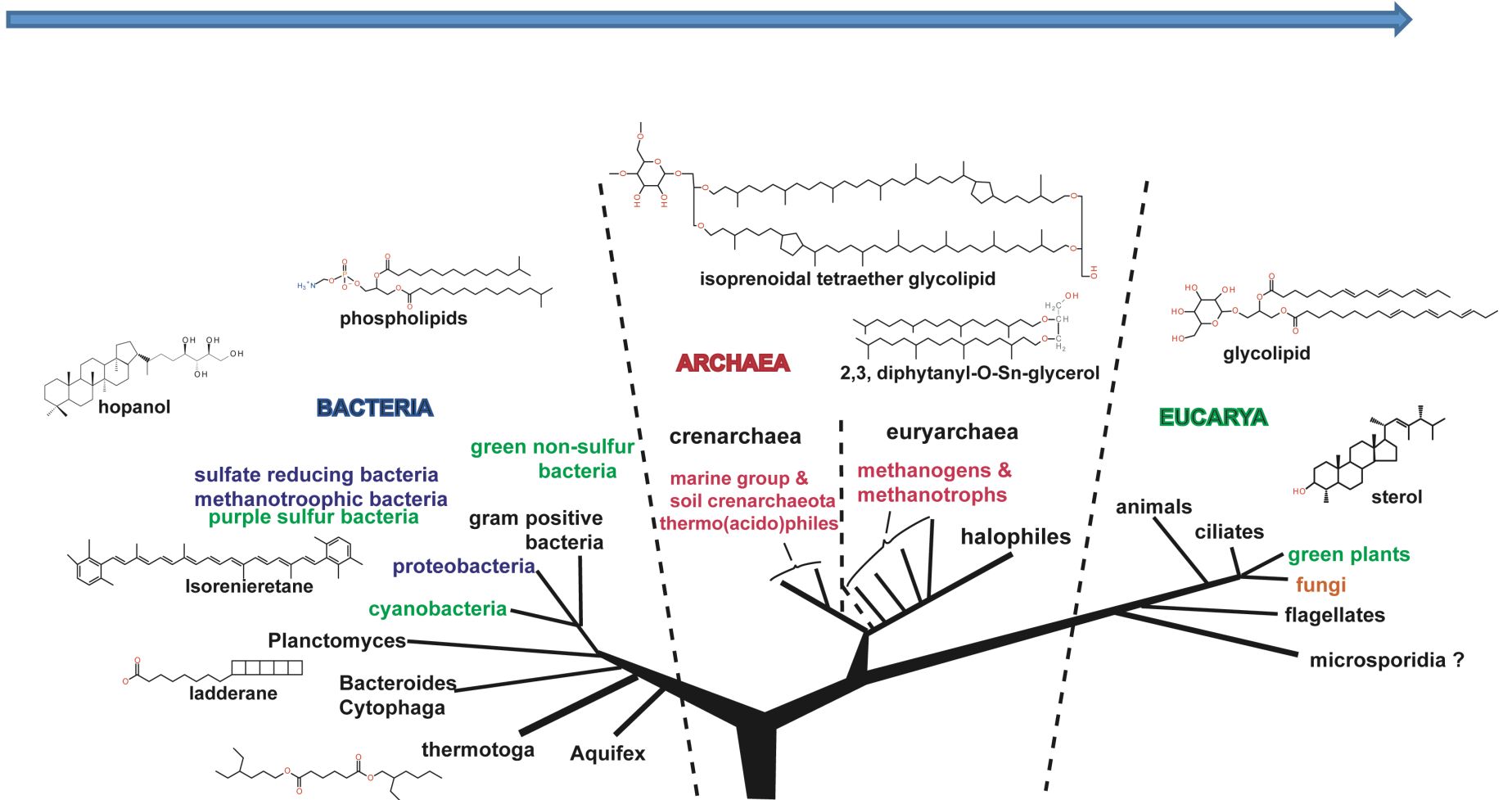
CSRA MAY HELP TO IMPROVE MODELS

What is a biomarker in environmental and climate science

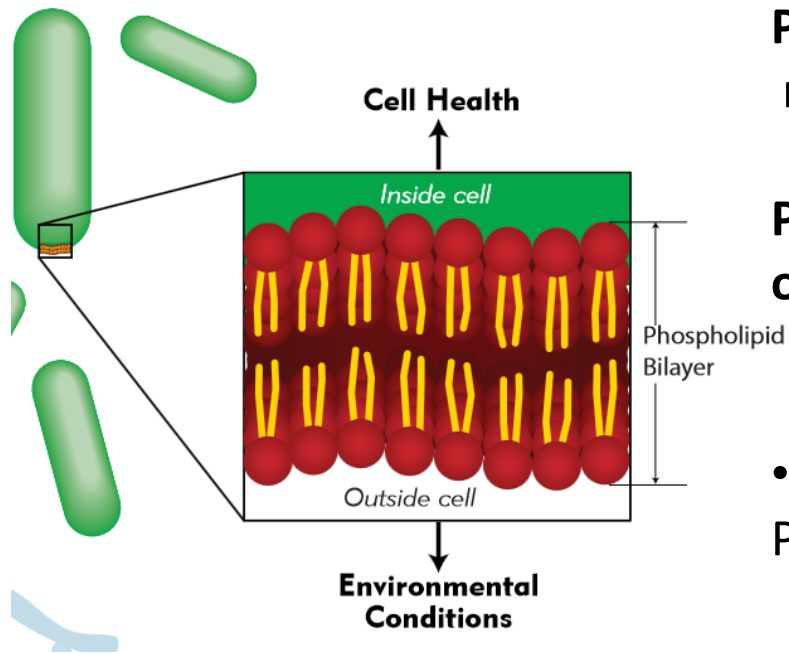
Molecular biological markers, or **biomarkers**, are natural products that can be traced to a particular biological origin.

Organic compounds with **specific biological sources**, whose structures can be **preserved** through geologic time.

Example of BIOMARKERS



What are PLFAs



Phospholipid fatty acids (PLFAs) are a main component of cell bacteria.

PLFAs analysis provides direct information on the entire microbial community.

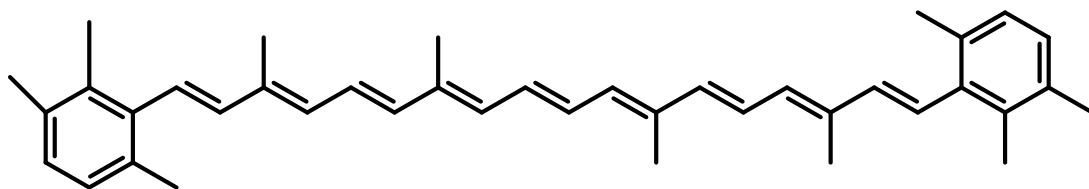
- Biomass
PLFAs represent all living cells.
- Population “Fingerprint”
Some organisms produce specific or signature types of PLFA biomarkers allowing quantification of important microbial functional groups (e.g. iron reducers, sulfate reducers, or fermenters).

PLFAs: as microbial population fingerprint



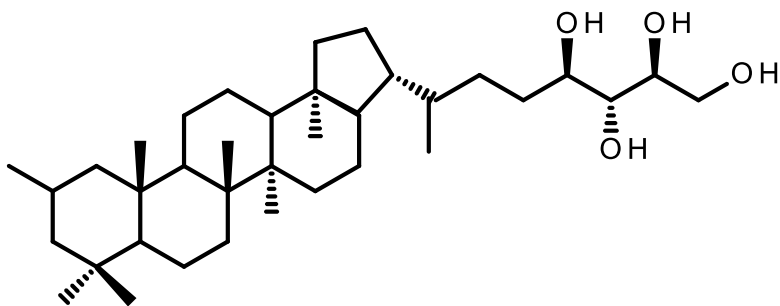
PLFAs Type	Bacterial group	Potential relevance
Monoenoic (Monos)	Gram-negative; Proteobacteria (aerobes and anaerobes)	e.g. Hydrocarbon utilizing or Nitrogen fixing bacteria
Terminally branched Saturated	Firmicutes and Bacteroides	Firmicutes: anaerobic fermenting bacteria
Branched monoenoic	in sulfate reducing bacteria and Planctomycetes	Desulfobacter
Mid-Chain Branched Saturated	In sulfate reducing bacteria and <i>Actinomyces</i>	Often associated with iron reducing bacteria
Polyenoic (Polys)	Found in eukaryotes Eukaryotic (fungi, algae, protozoa, plants and animals)	Eukaryotic scavengers often prey on contaminant utilizing bacteria

Some more specific bacteria lipids



isorenieratene

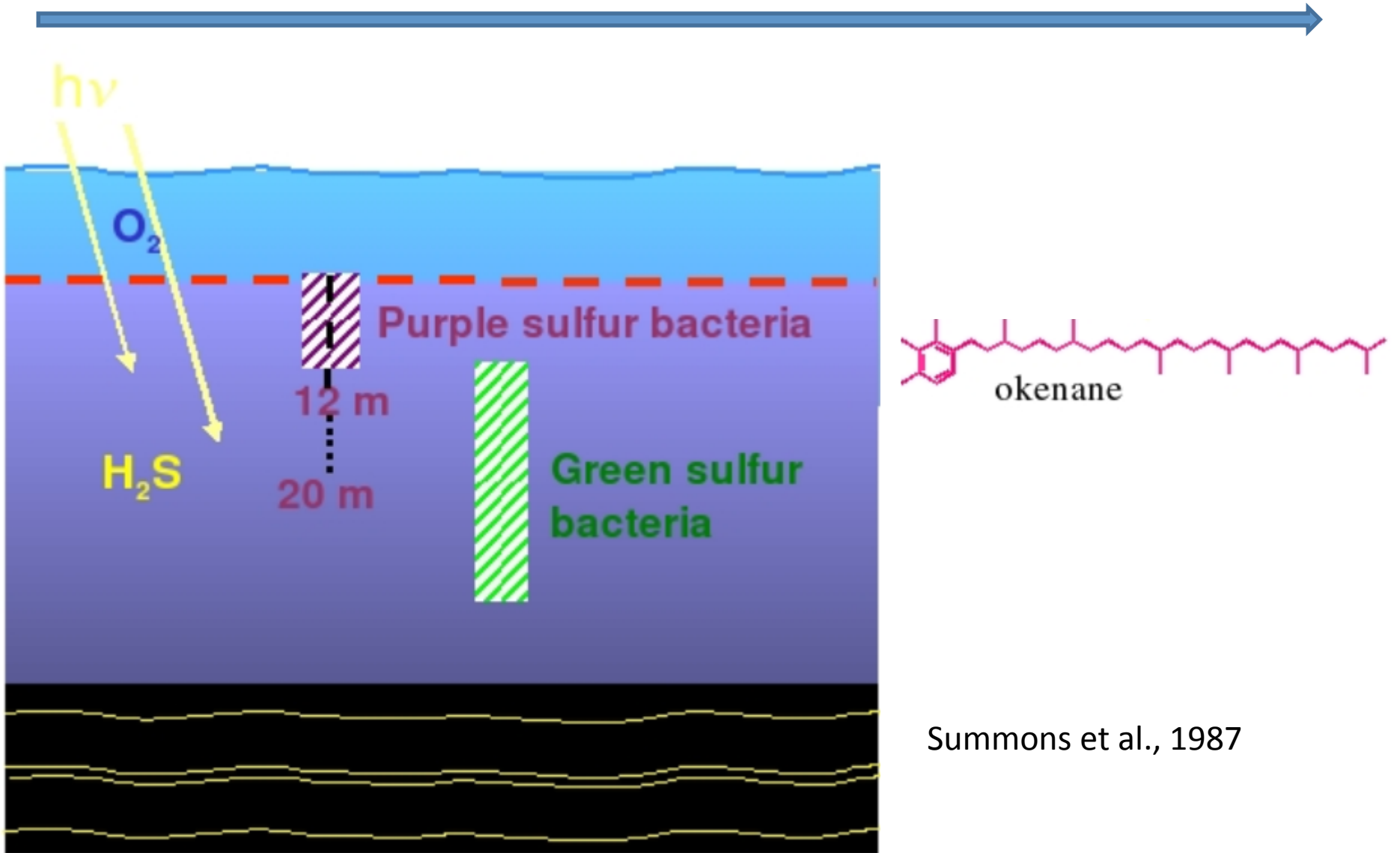
Green purple sulfur bacteria



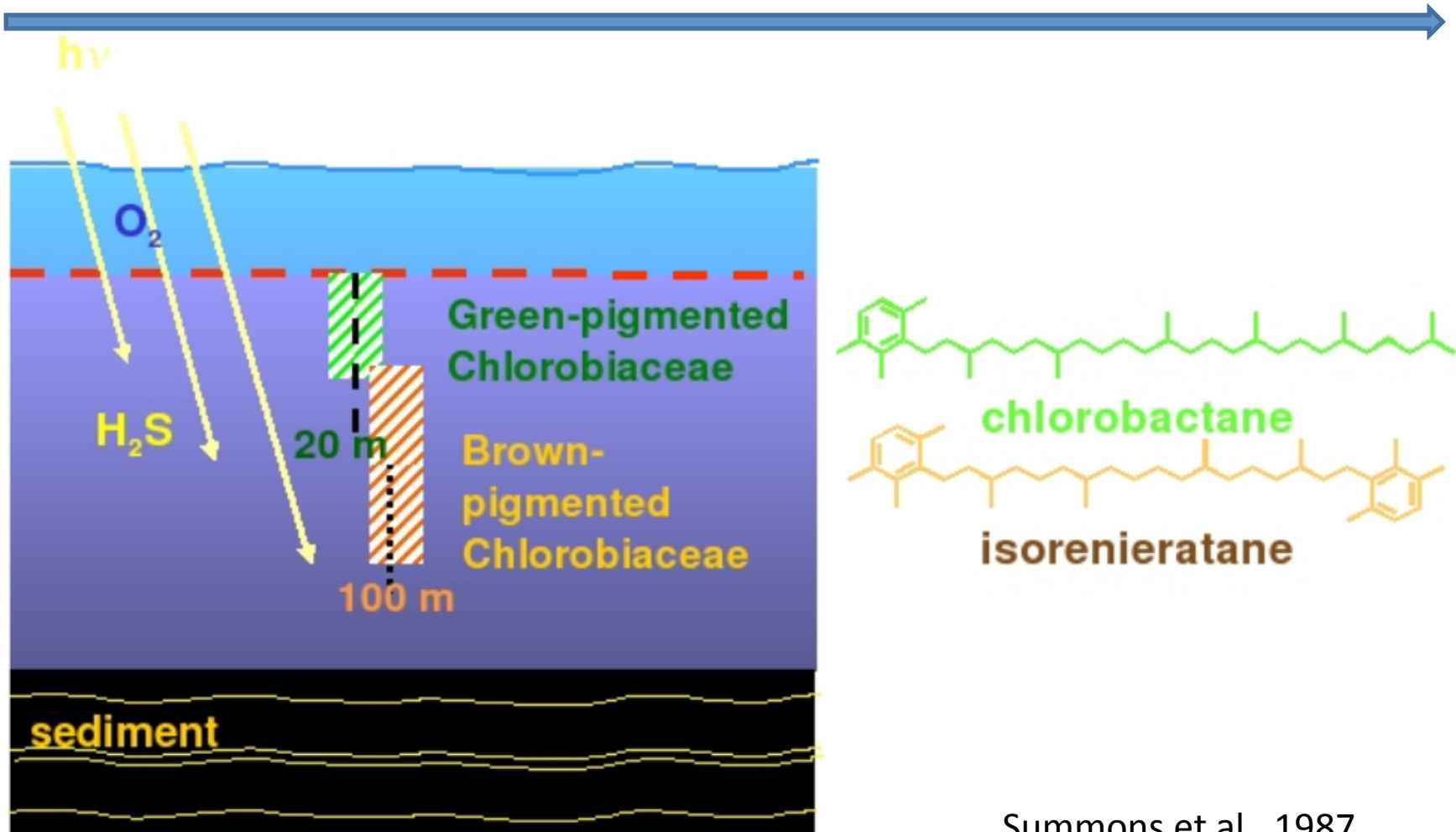
2-methyl hopanol

cyanobacteria

Some more specific bacteria lipids



Some more specific “bacteria” lipids

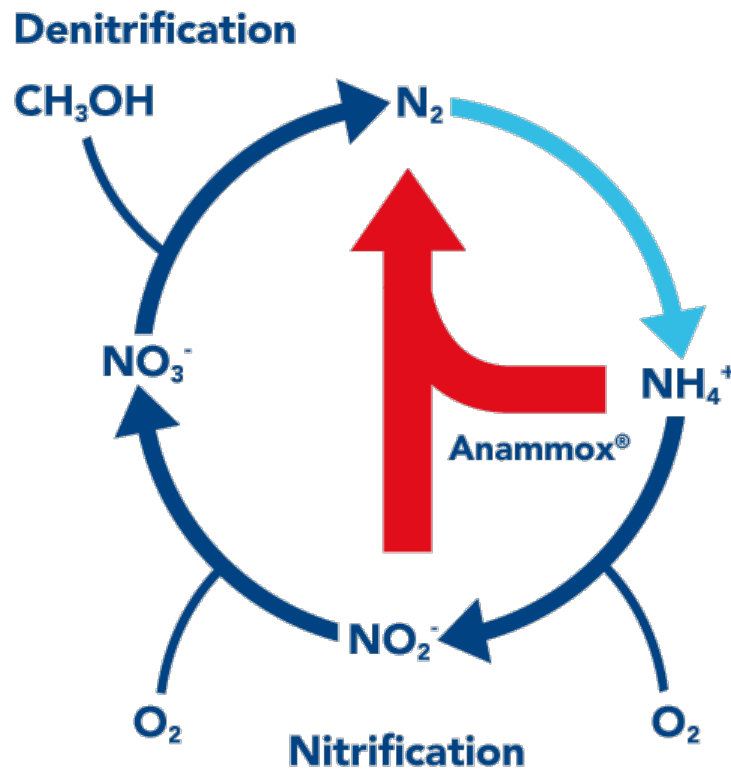


Summons et al., 1987

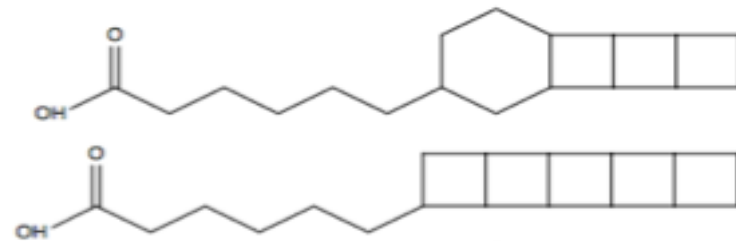
Some more specific bacteria lipids

Ladderanes (PLFAs) derived from anammox bacteria, an abbreviation for ANaerobic AMMonium OXidation

Nitrogen Cycle

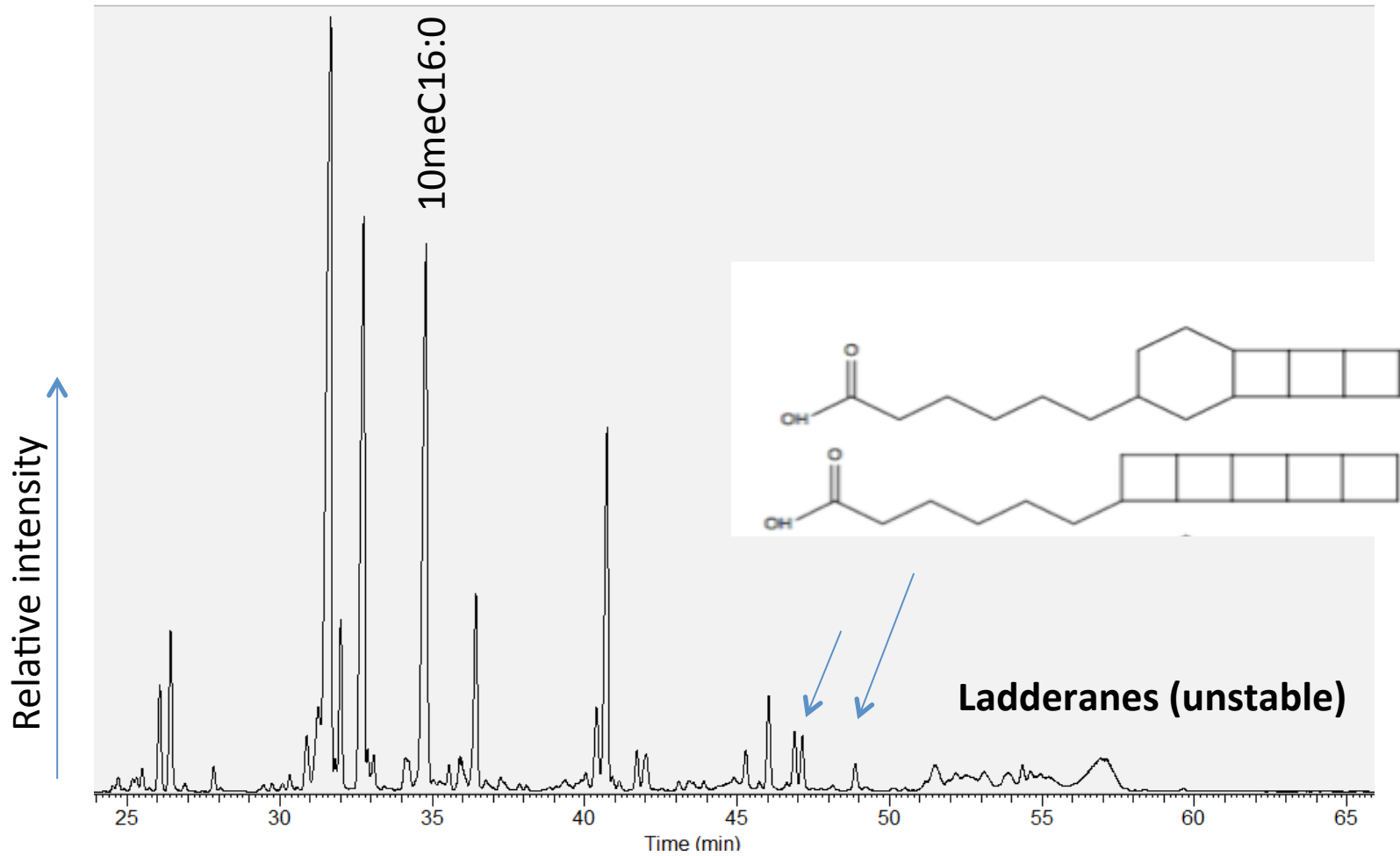


Ladderanes

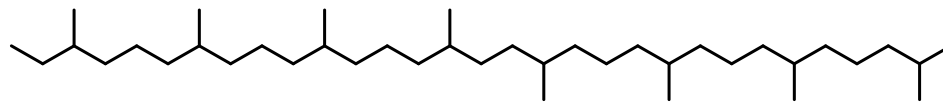


They have been proposed to be responsible of up to 70% of oceanic N_2 production, representing a major N sink.

Example of PLFAs profile (groundwater)

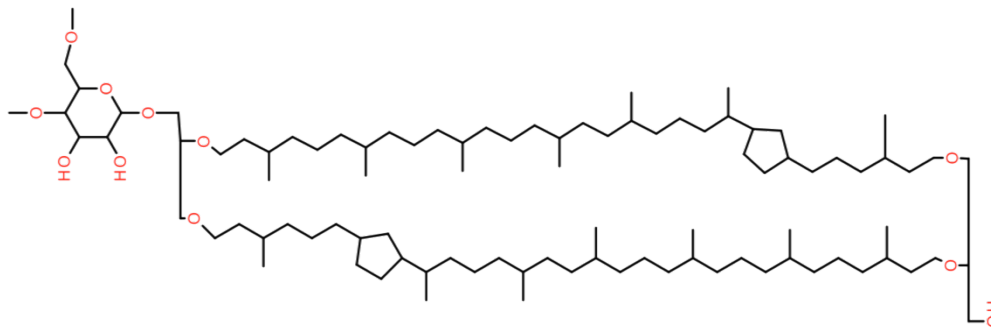


Some more specific archaea lipids



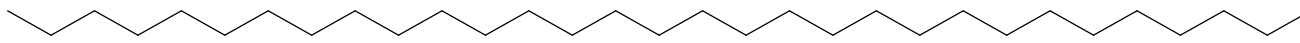
biphytane

Methanotrophic archaea



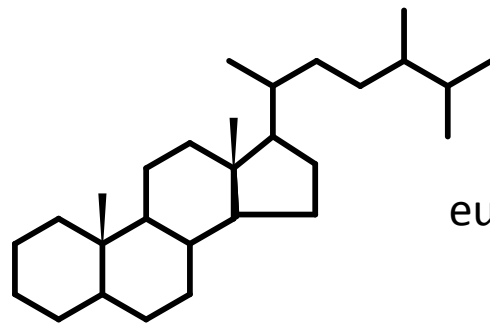
Glycerol dibiphytanyl glycerol *tetraether* (GDGT)

More specific prokaryote lipids

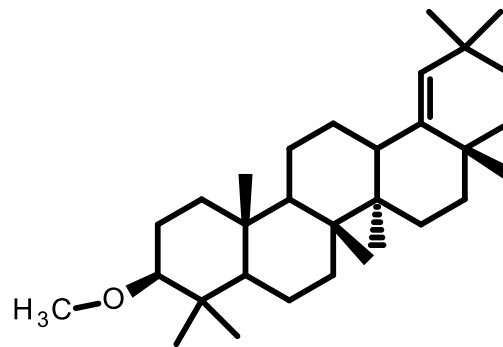


Long chain *n*-alkanes

Terrestrial plants



eucaryotes

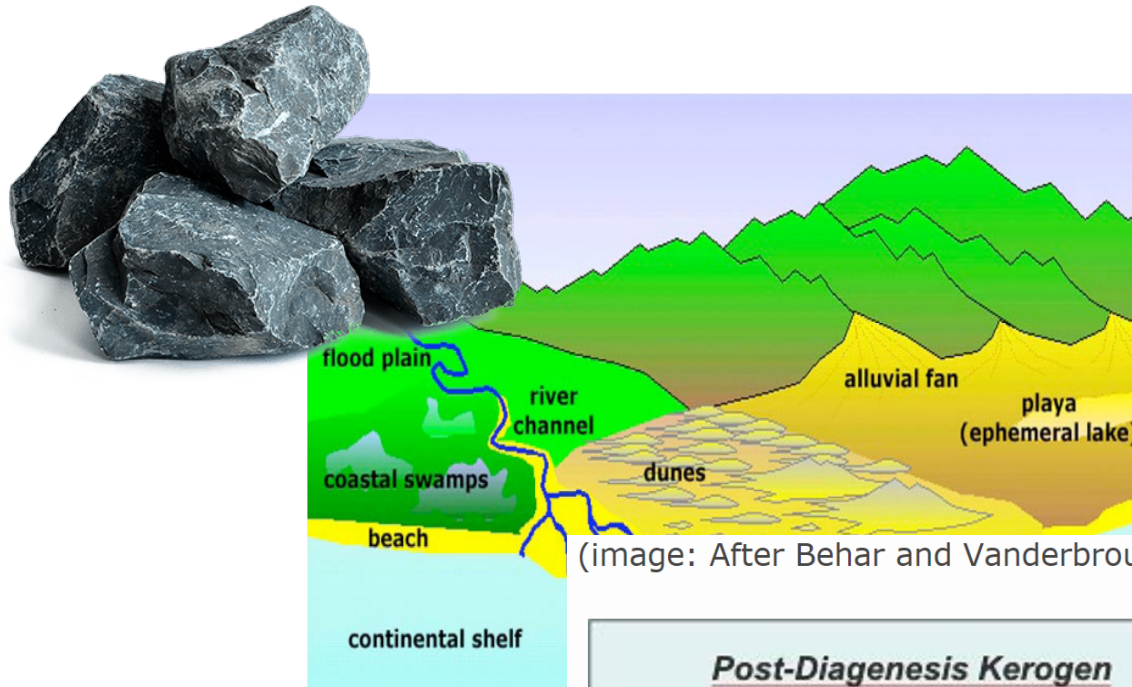


miliacine

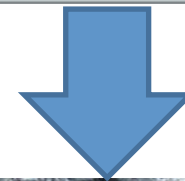
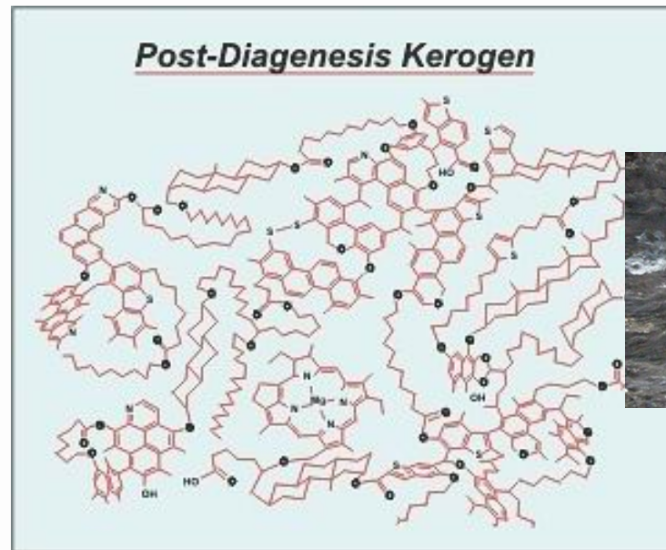
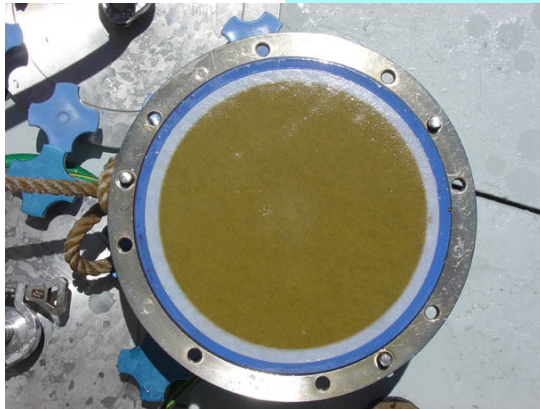
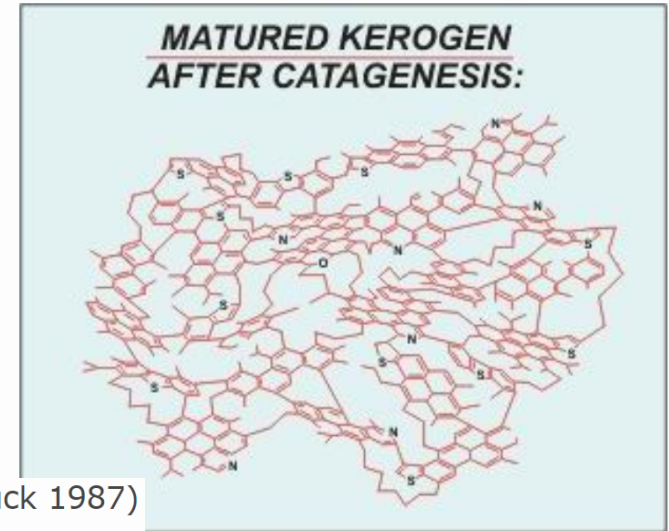
Formation and origin of biomarkers



(image: After Behar and Vanderbrouck 1987)



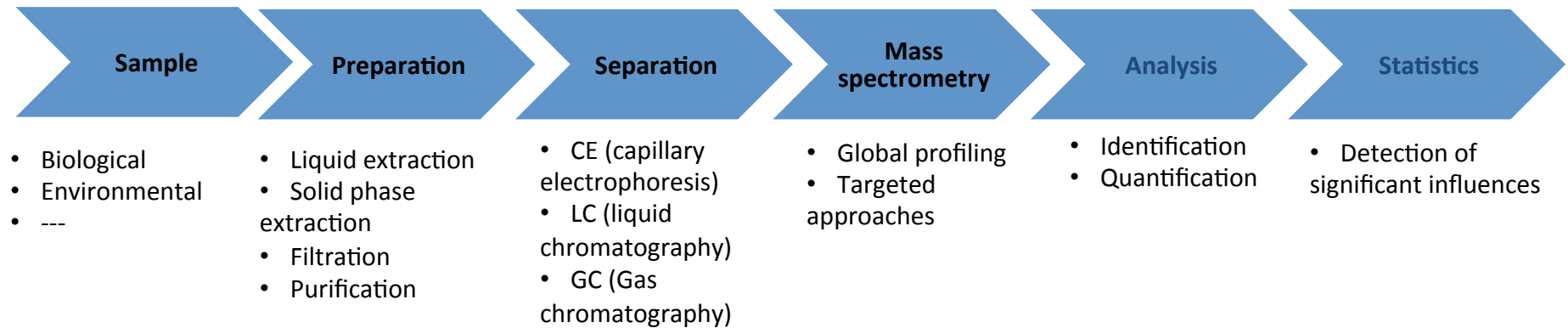
(image: After Behar and Vanderbrouck 1987)



Free lipids



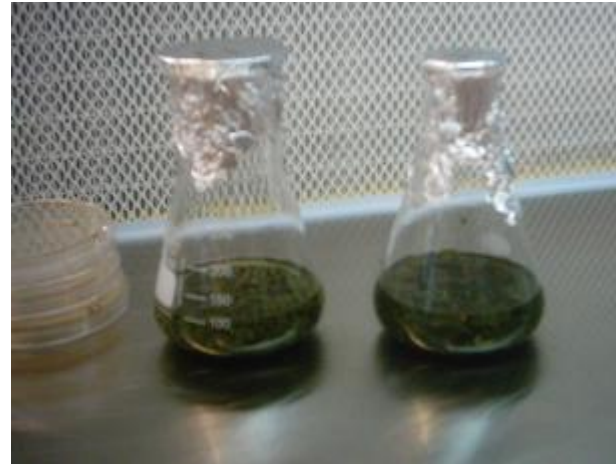
General workflow



Lipid extract



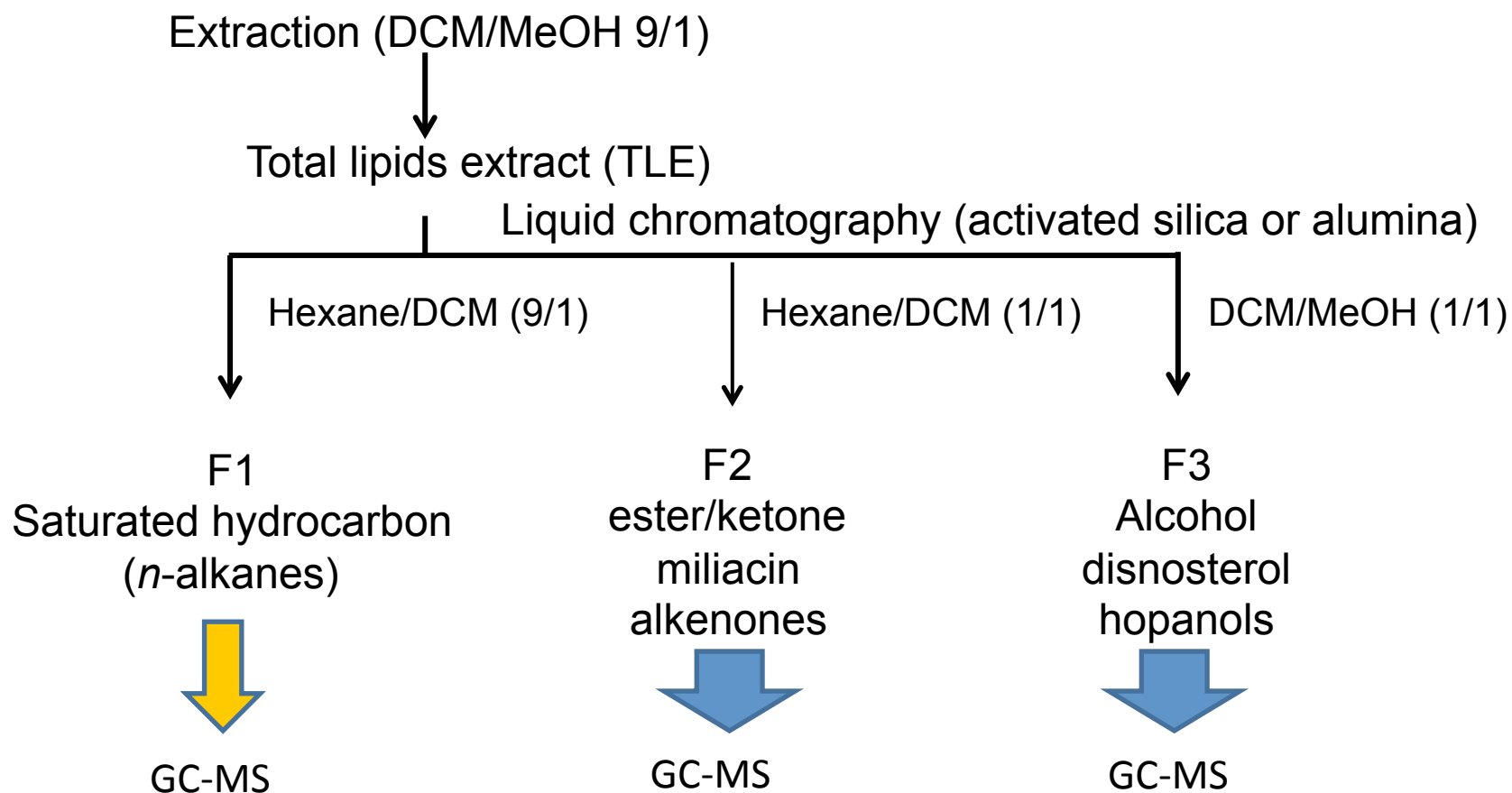
Mature samples



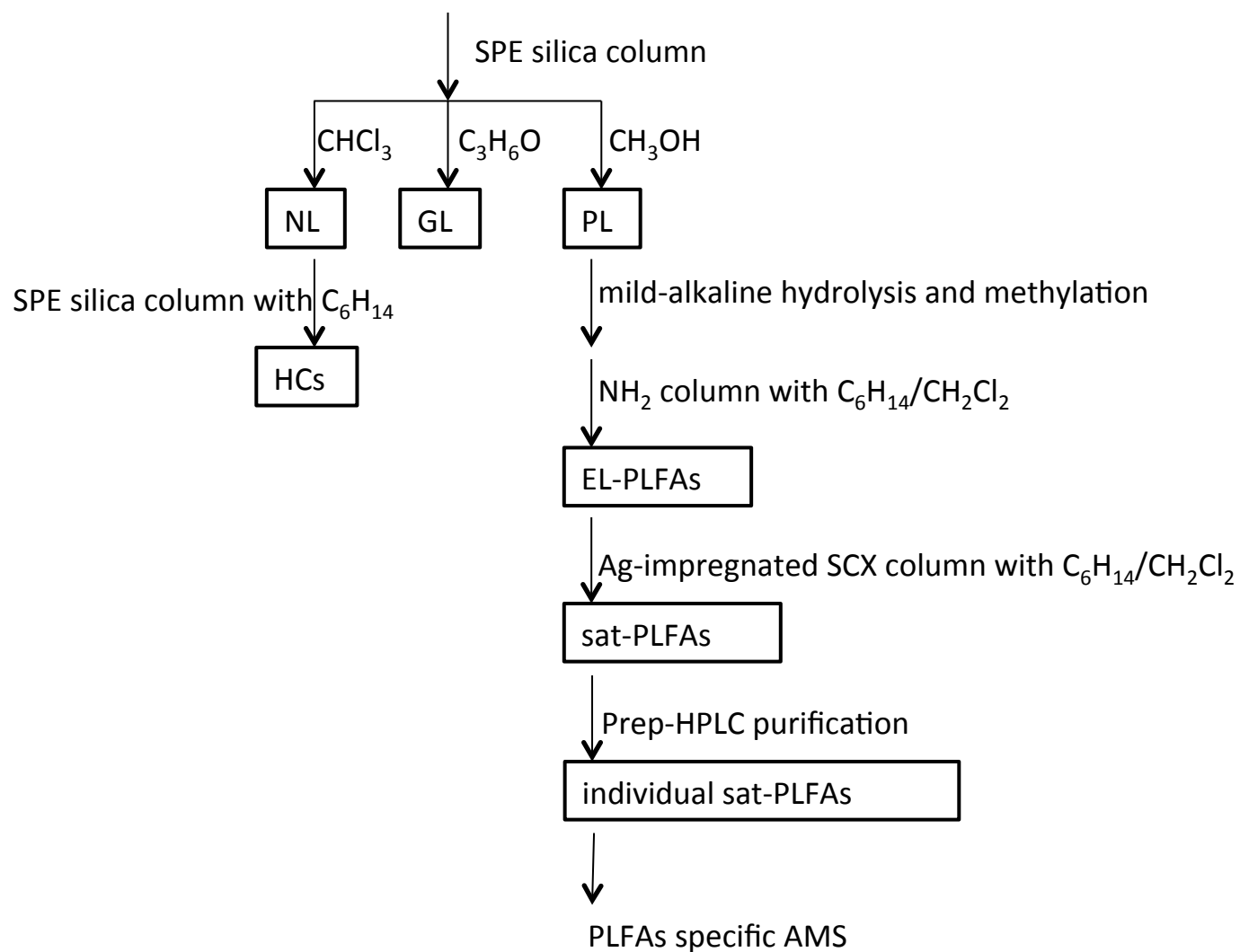
Plant extract



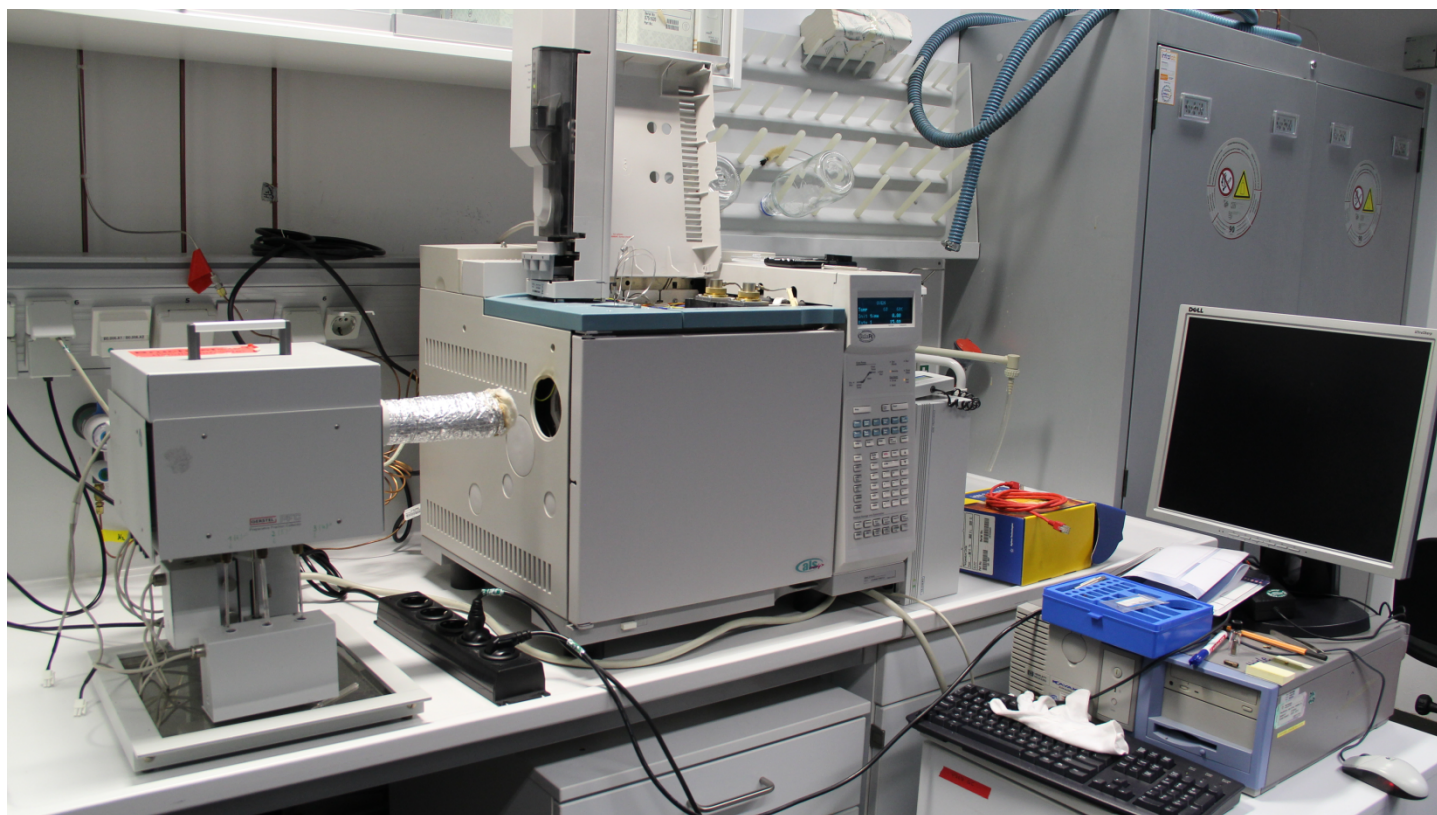
Typical purification of mature samples



Typical purification of fresh samples



Compound purification –prep –GC

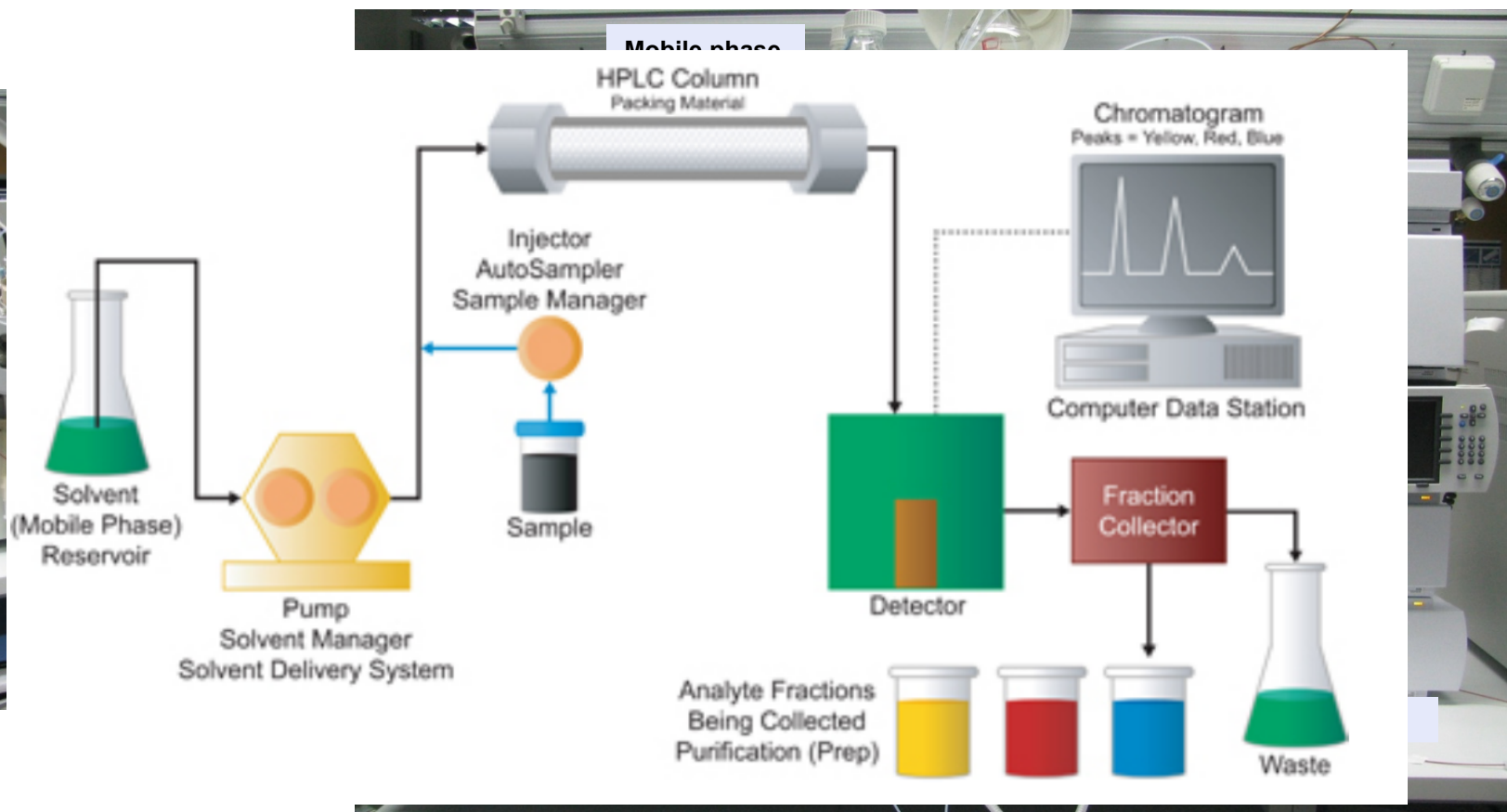


Compound purification –prep – HPLC

HPLC-MS (quadrupole)



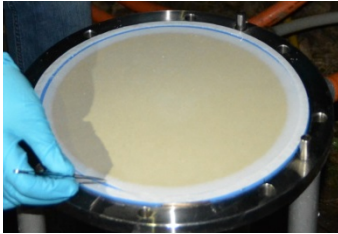
MS (ESI)



HPLC

Chemical treatment

Samples are frozen
(-80°C) until analyze



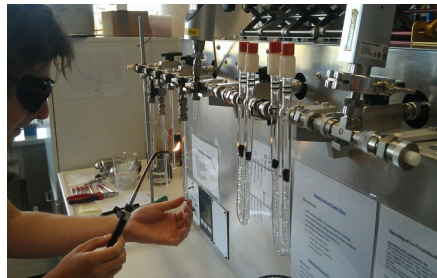
Lipid extraction



Chemical pre-
purification



Extraction lines



Prep-HPLC



GC-TQMS



Lipids identification

AMS



Lipids isolation

$\Delta^{14}\text{C}$ PLFAs

Comparison between HPLC and GC



Sample volatility

HPLC

No volatility requirement

Sample must be soluble
in mobile phase

GC

Sample must be
volatile

Sample Polarity

HPLC

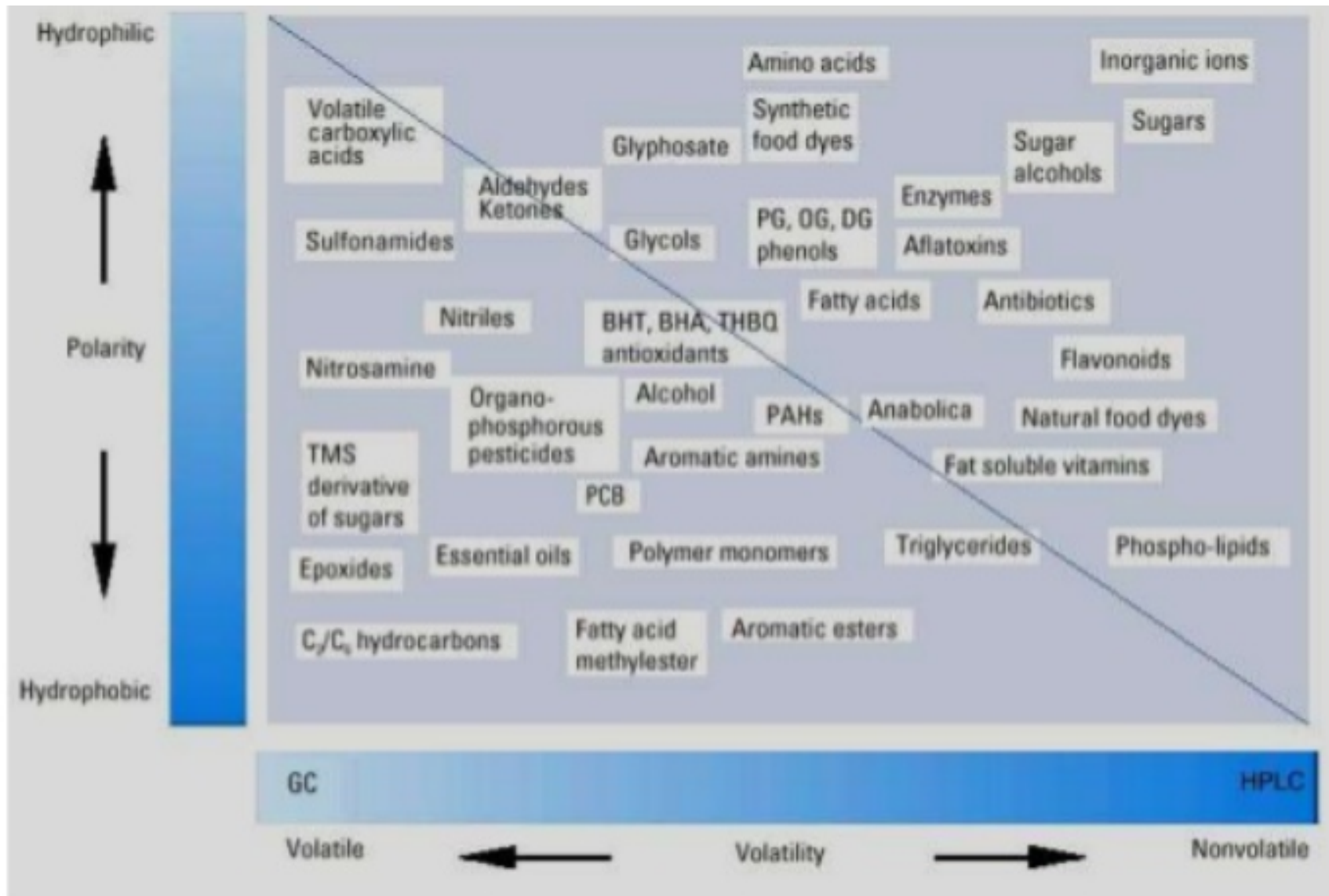
Separates both polar and
non polar compounds

PAH-inorganic ions

GC

Separates both polar and
non polar compounds

Comparison between HPLC and GC



Comparison between HPLC and GC



Sample thermal lability

HPLC

Close to room temperature

GC

Sample must be able to survive high temp. injection

Sample molecular weight

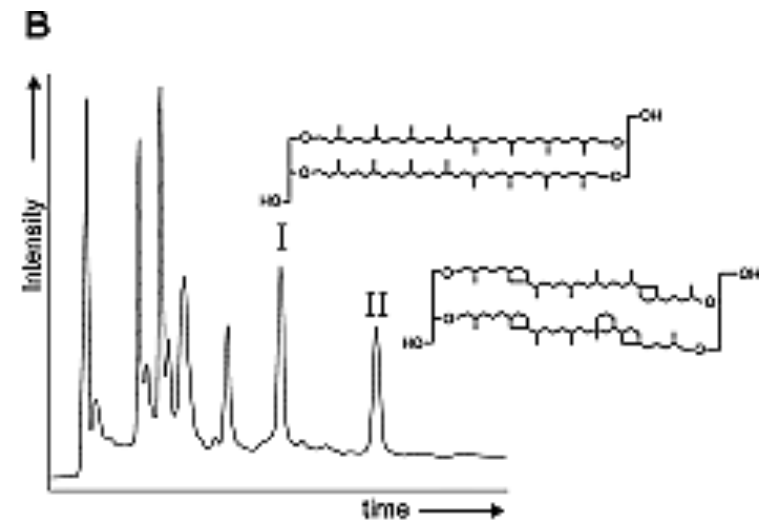
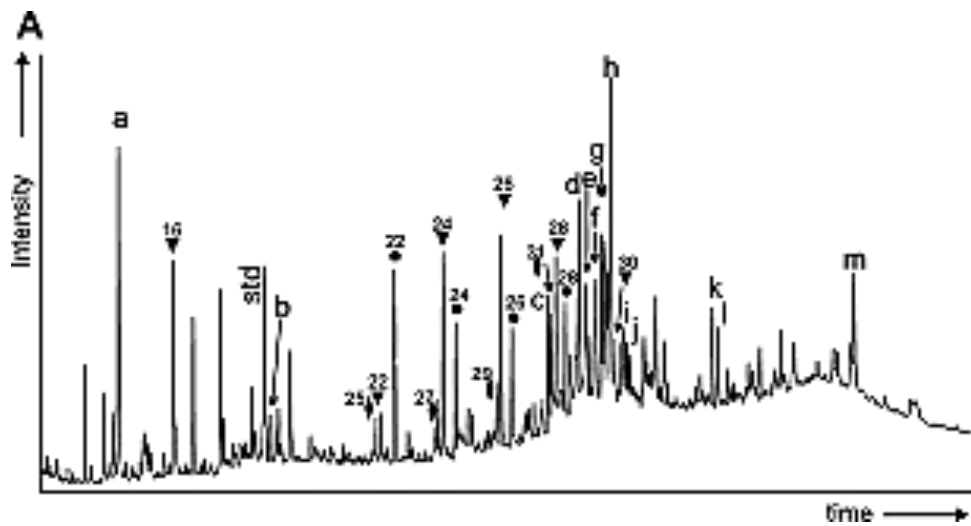
HPLC

No theoretical upper limit

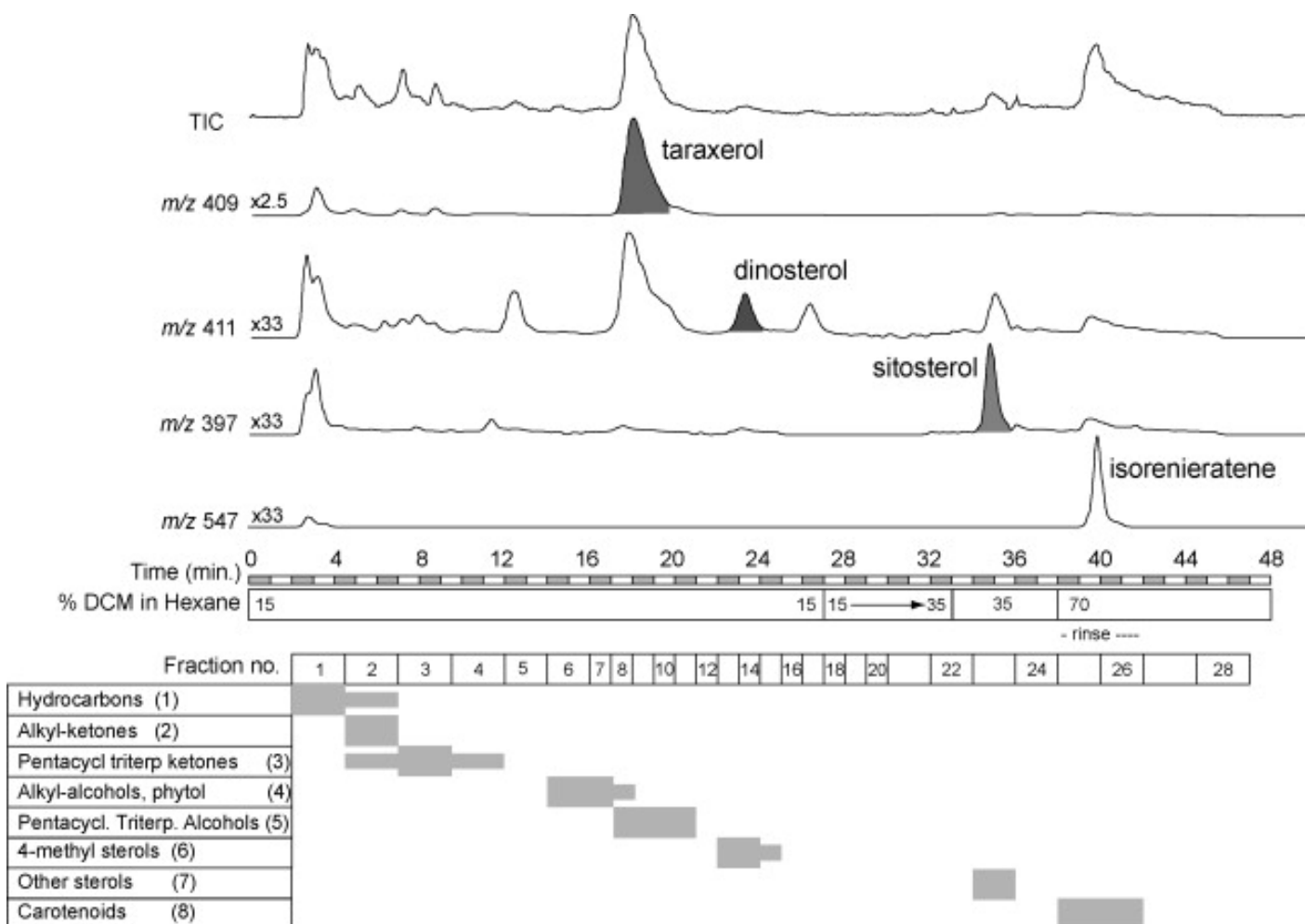
GC

Typically < 500 amu

Example of sterol fraction

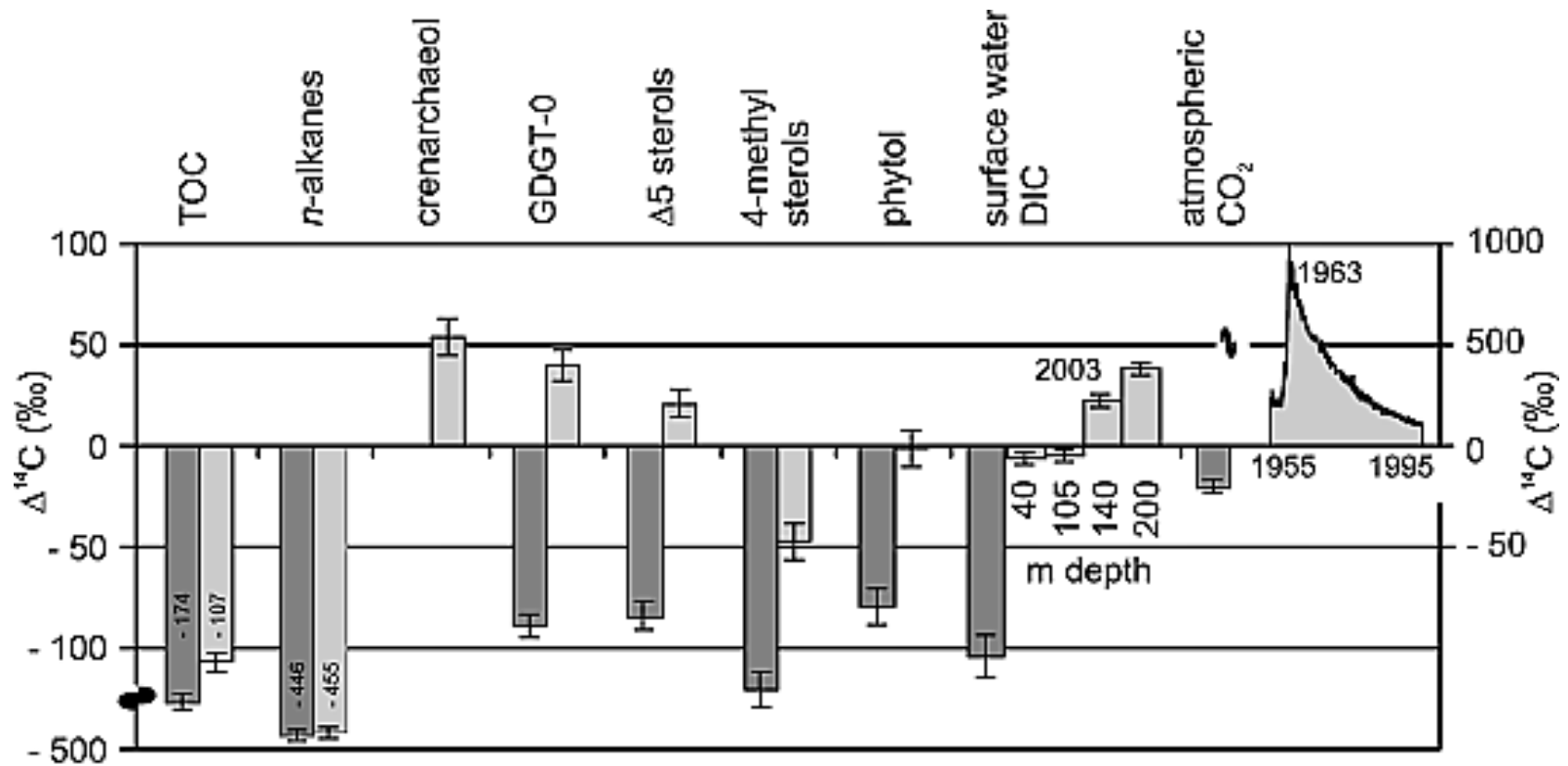


Example of sterol fraction run on HPLC



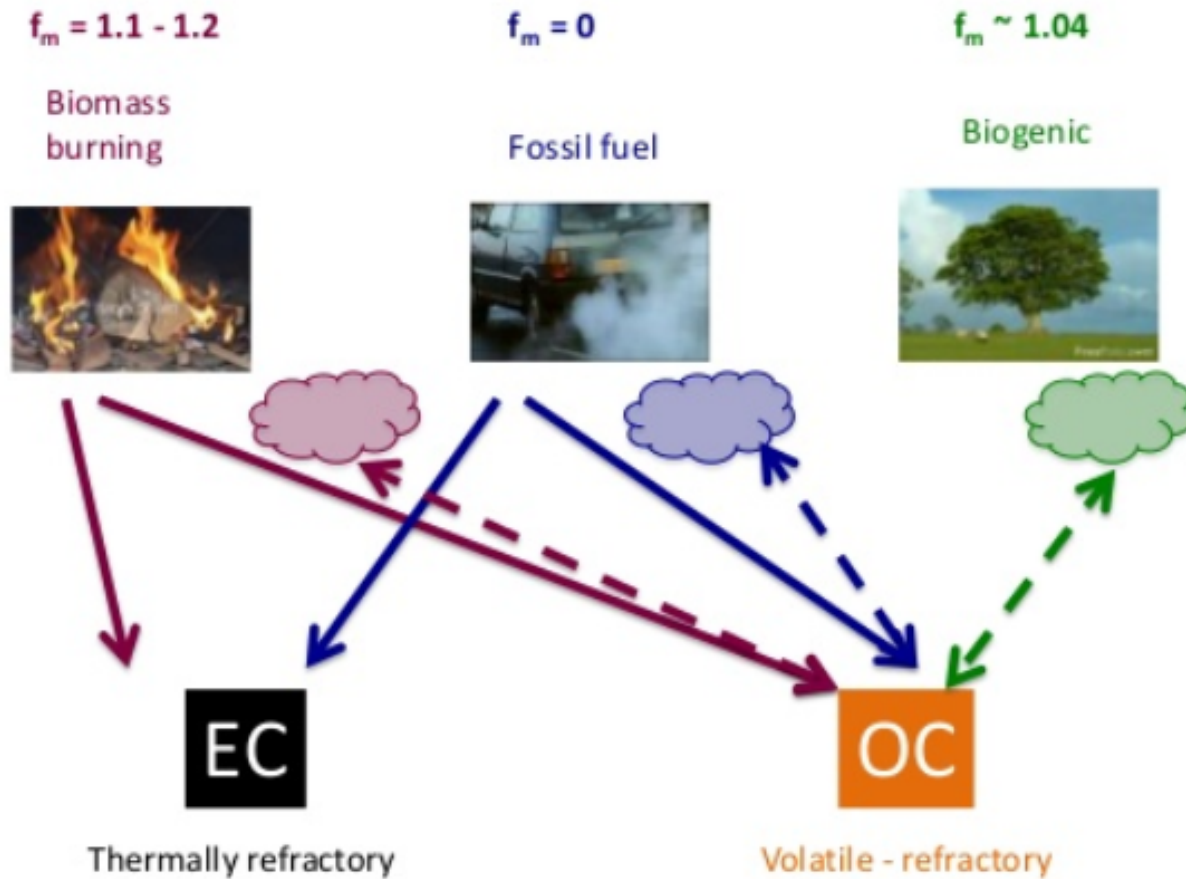
Smittenberg et al., 2007, Journal of chromatography A

Example of sterol fraction run on HPLC for "CSRA"

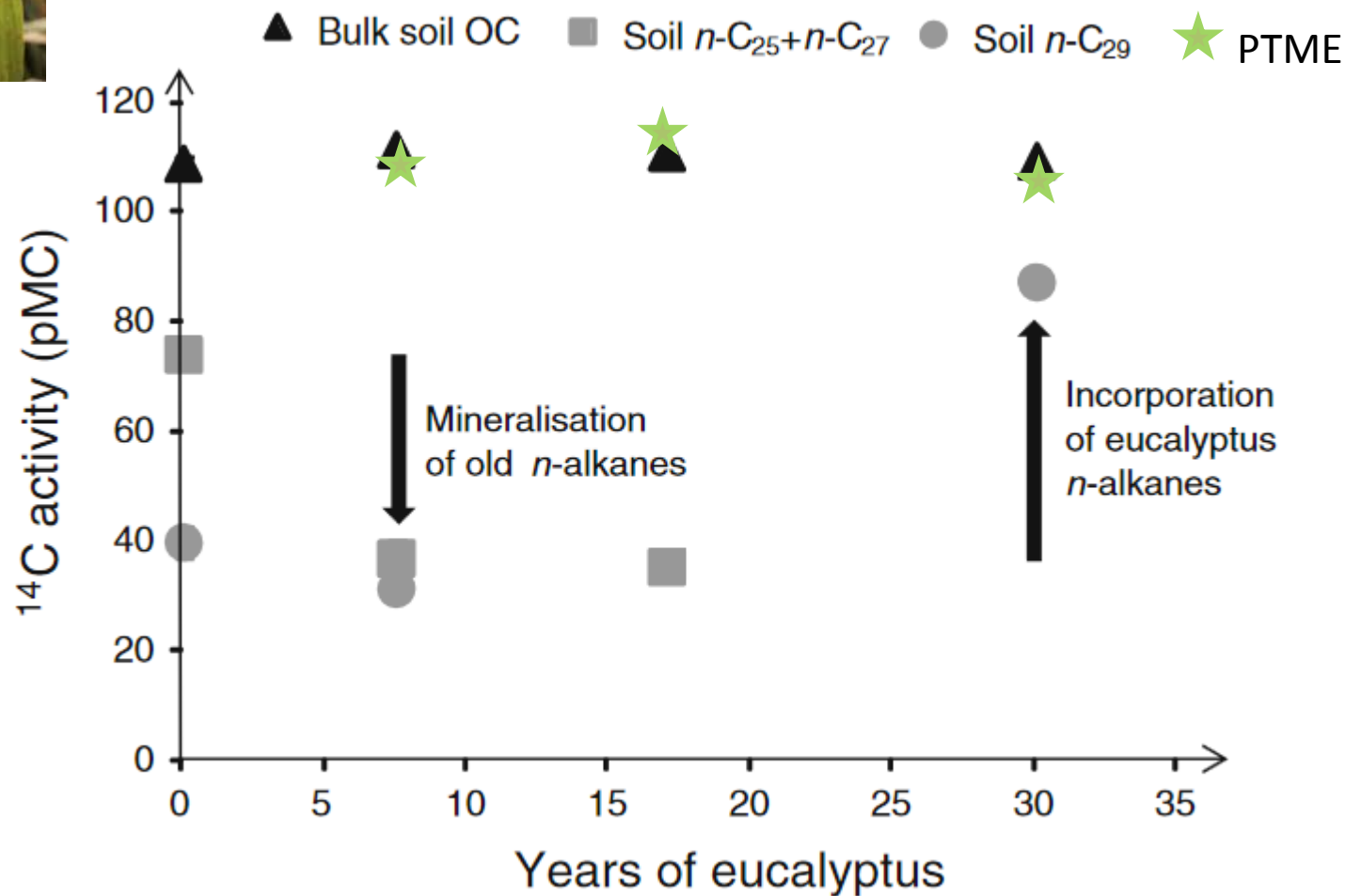
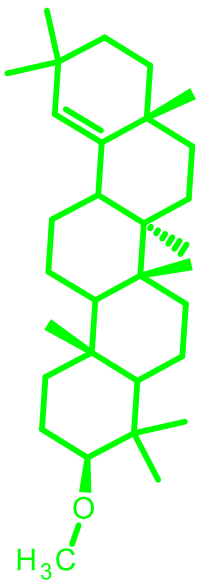


Smittenberg et al., 2006 Paleoceanography

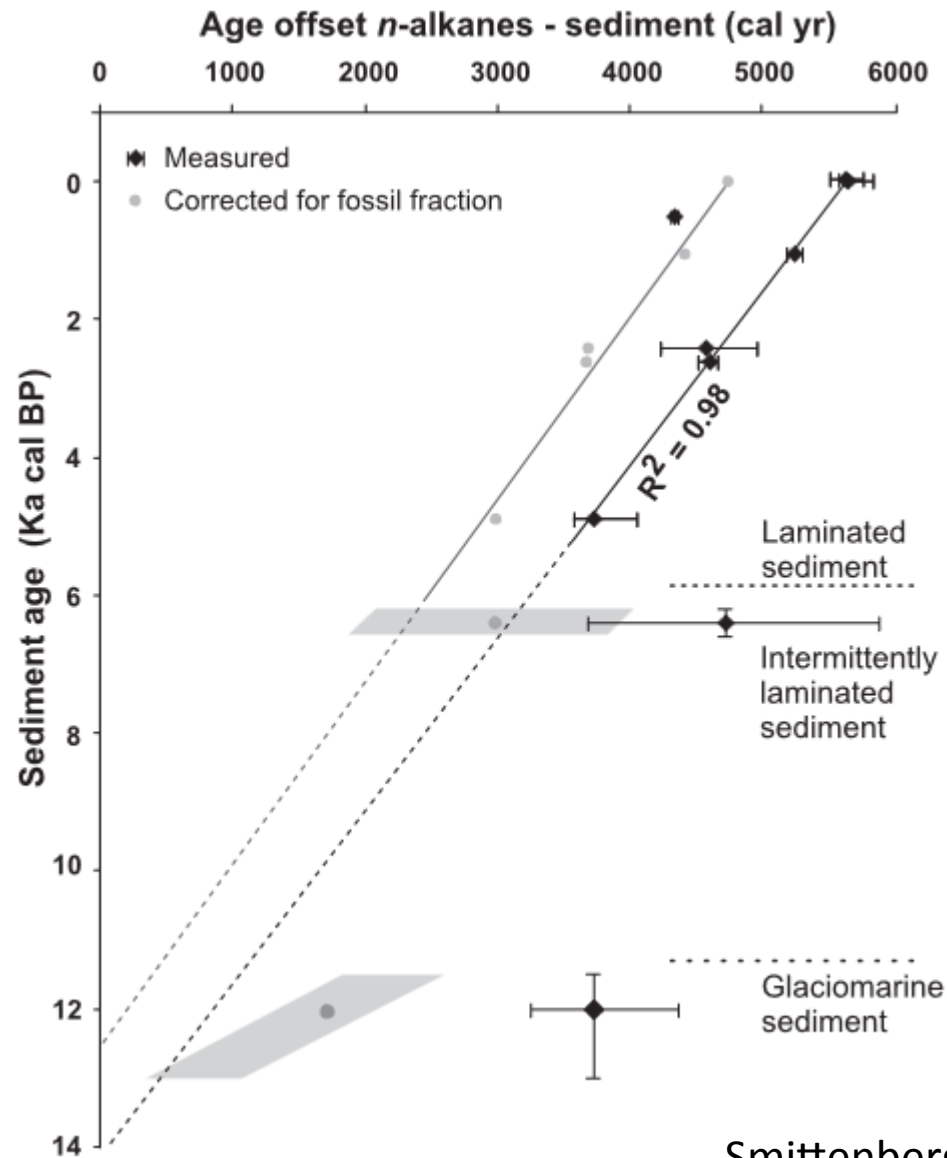
CSRA of aerosol used for source appointment



Different compound stabilization in savanna soil



Stabilization of wax *n*-alkane in sediments



Smittenberg et al., 2006 Science

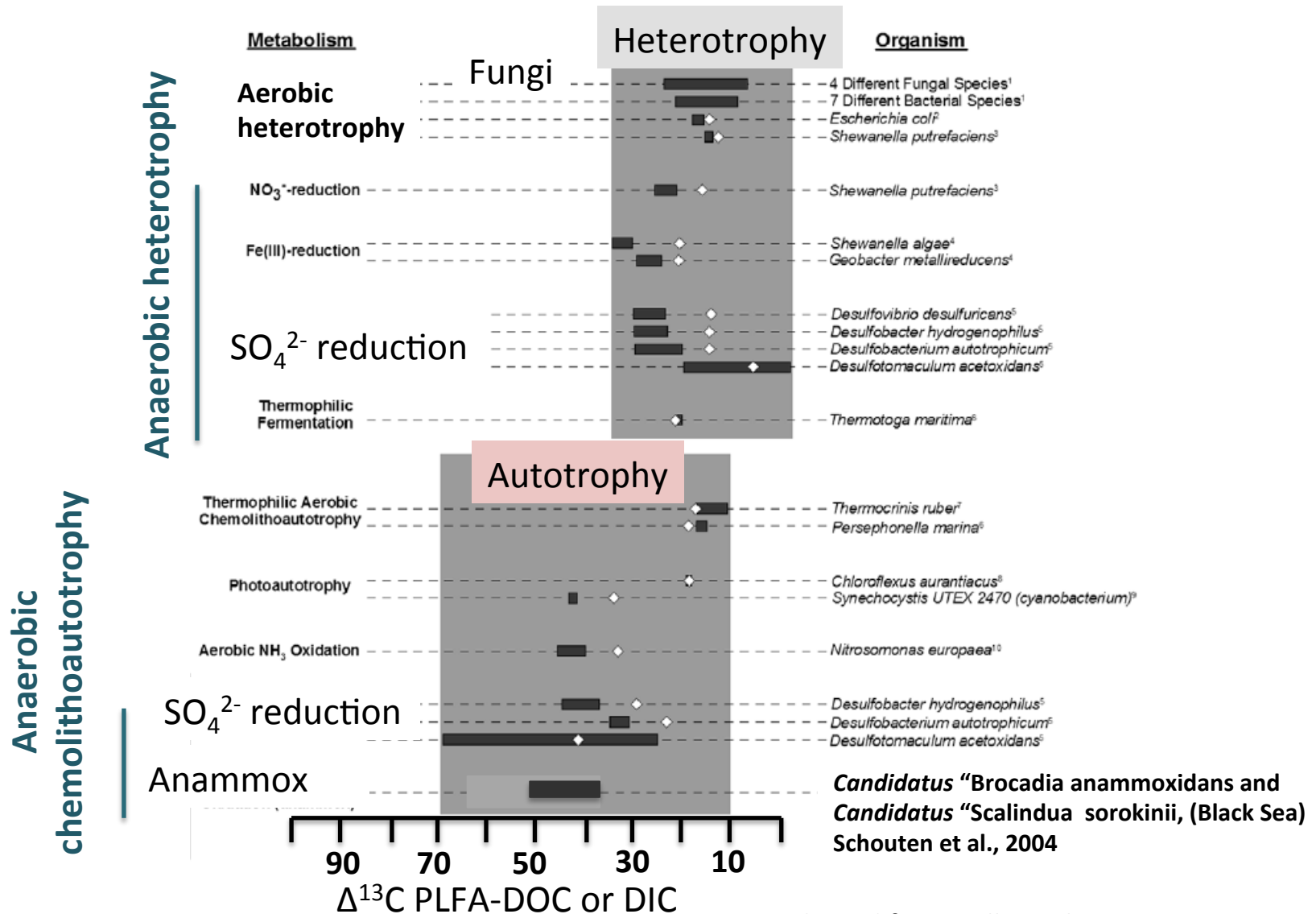
A EXAMPLE OF PLFA CSRA in groundwater



to help to define

- **the source of carbon used by an organism and the apparent age of the reservoir providing this source.**
- **the metabolic pathways of organisms**

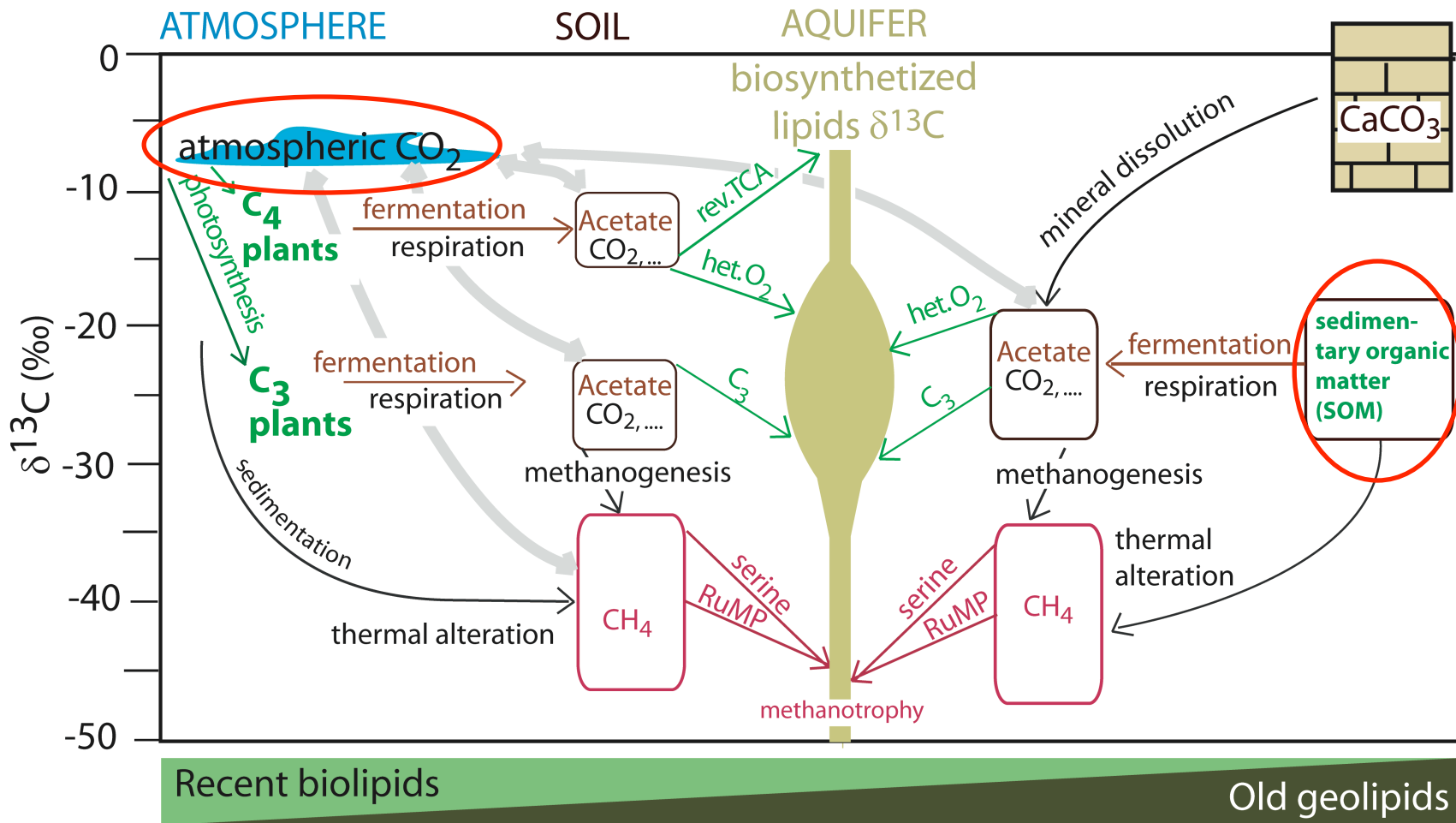
PLFAs $\delta^{13}\text{C}$ values have a lot of overlap



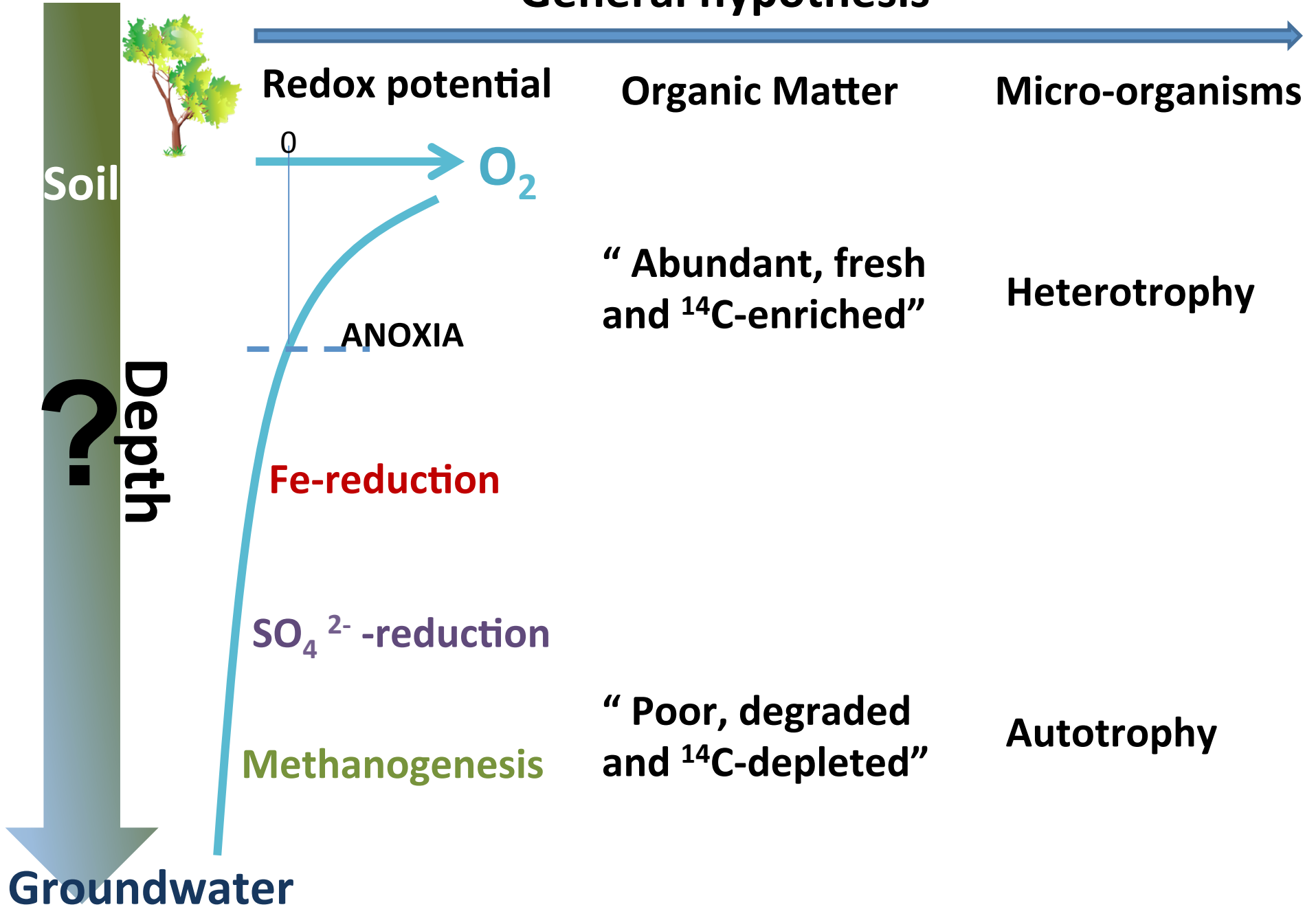
Adapted from Mills et al., 2010

PLFAs ^{14}C values help to definite the C source

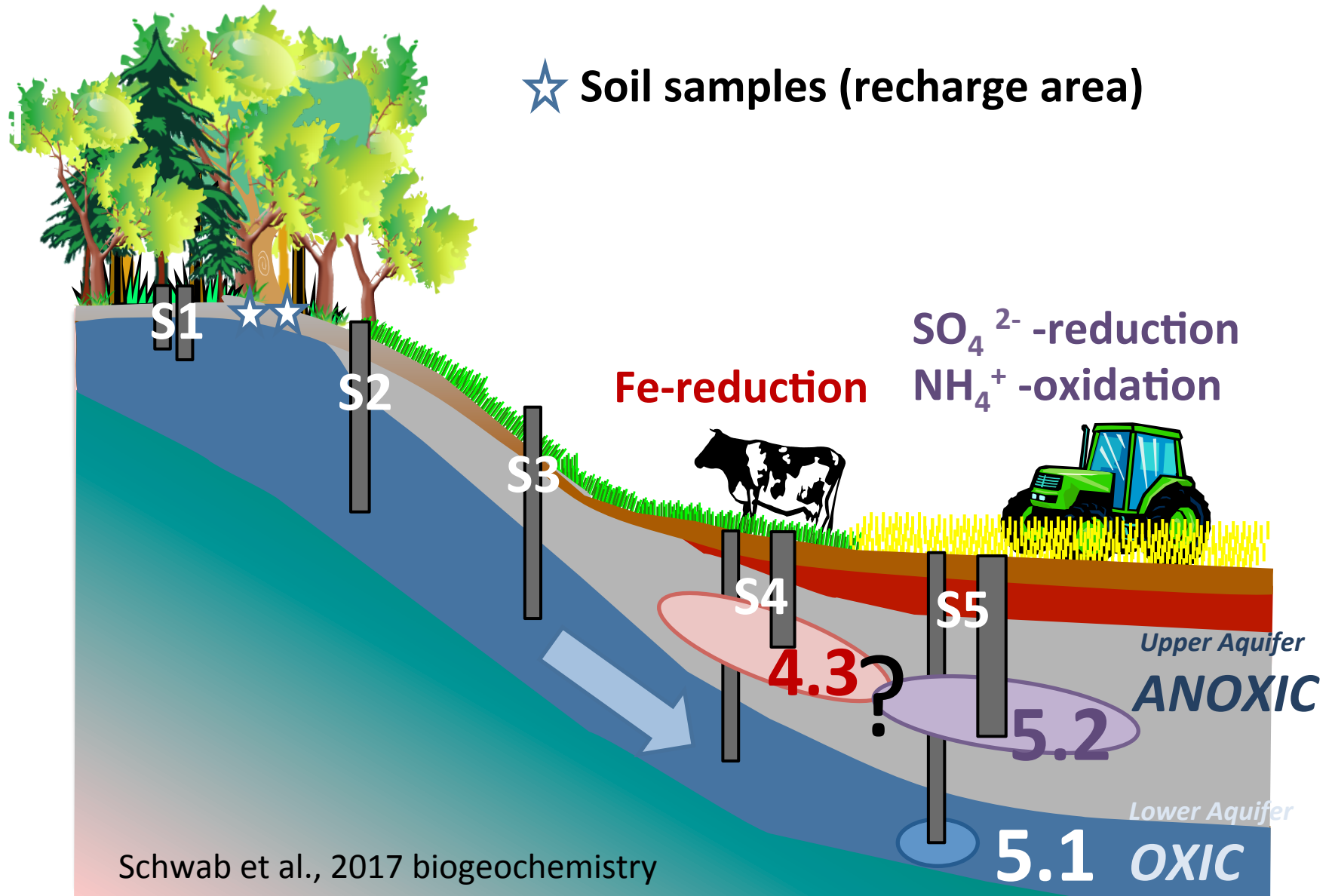
Mainly 3 organic pools in groundwater



General hypothesis



Condition of the sampling sites



Sampling



**View borehole
(ca. 10'000L)**



**Filtration system
pre-combusted (5h 500°C)
glass fiber membrane
Ø 293 mm, 0.3 µm**

Sampling

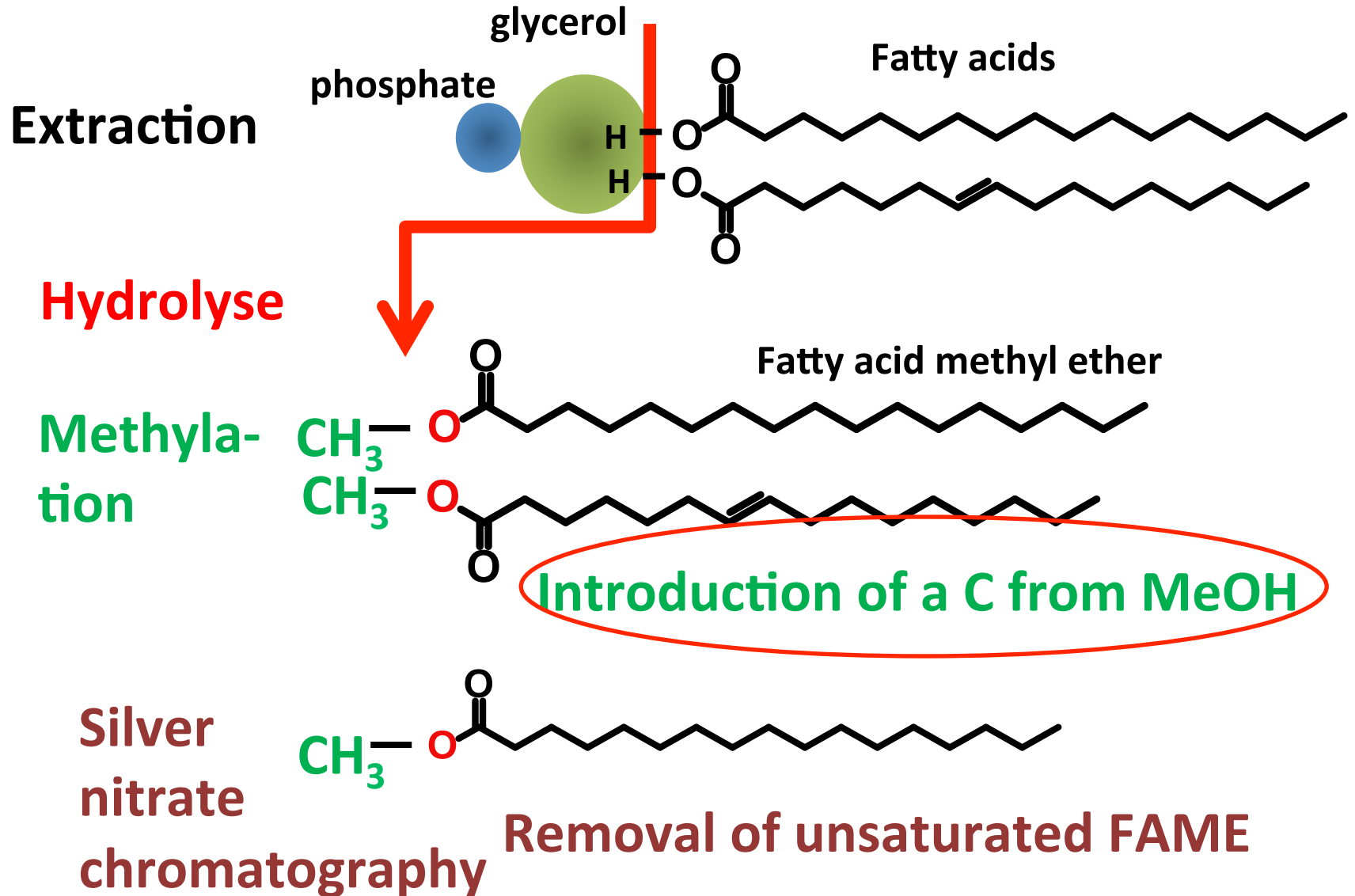


**H5.2 sulfate-rich well
(ca. 10'000L)**



**H4.3 Iron-rich well
(ca. 10'000L)**

Chemical treatment



Semi pre-HPLC

Eluting peaks in a C₁₈ HPLC column

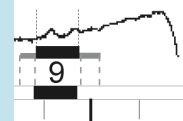
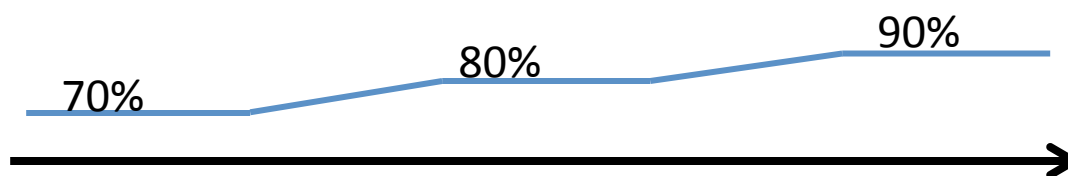
(150 x 4.6 mm, 3 μm)

Facts:

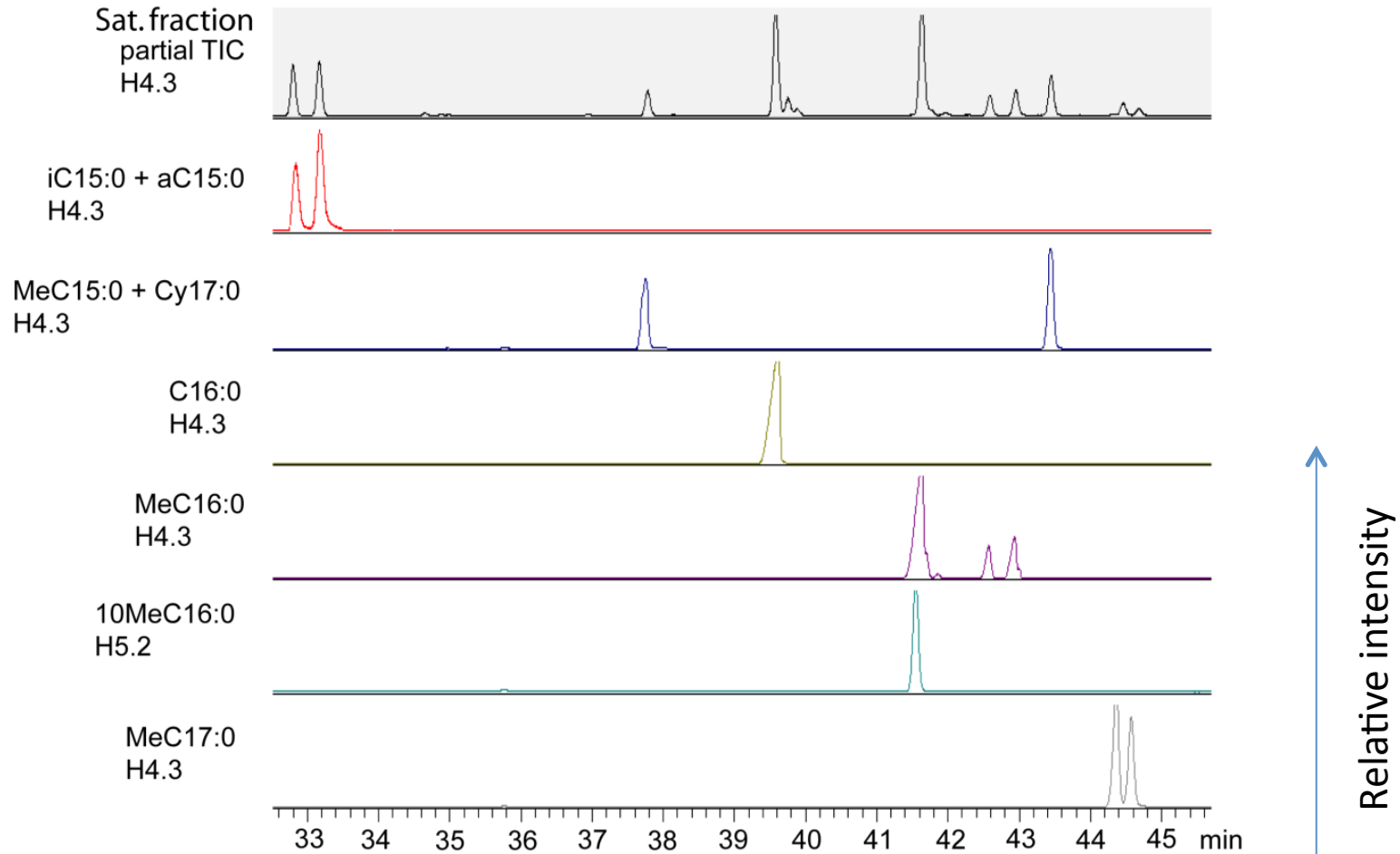
- Between 5 up to 10 μg of a single compound per injection
- Need between 15 and 10 injections for collecting ca. 100 μg
- Can collected many different compounds per injection

compound

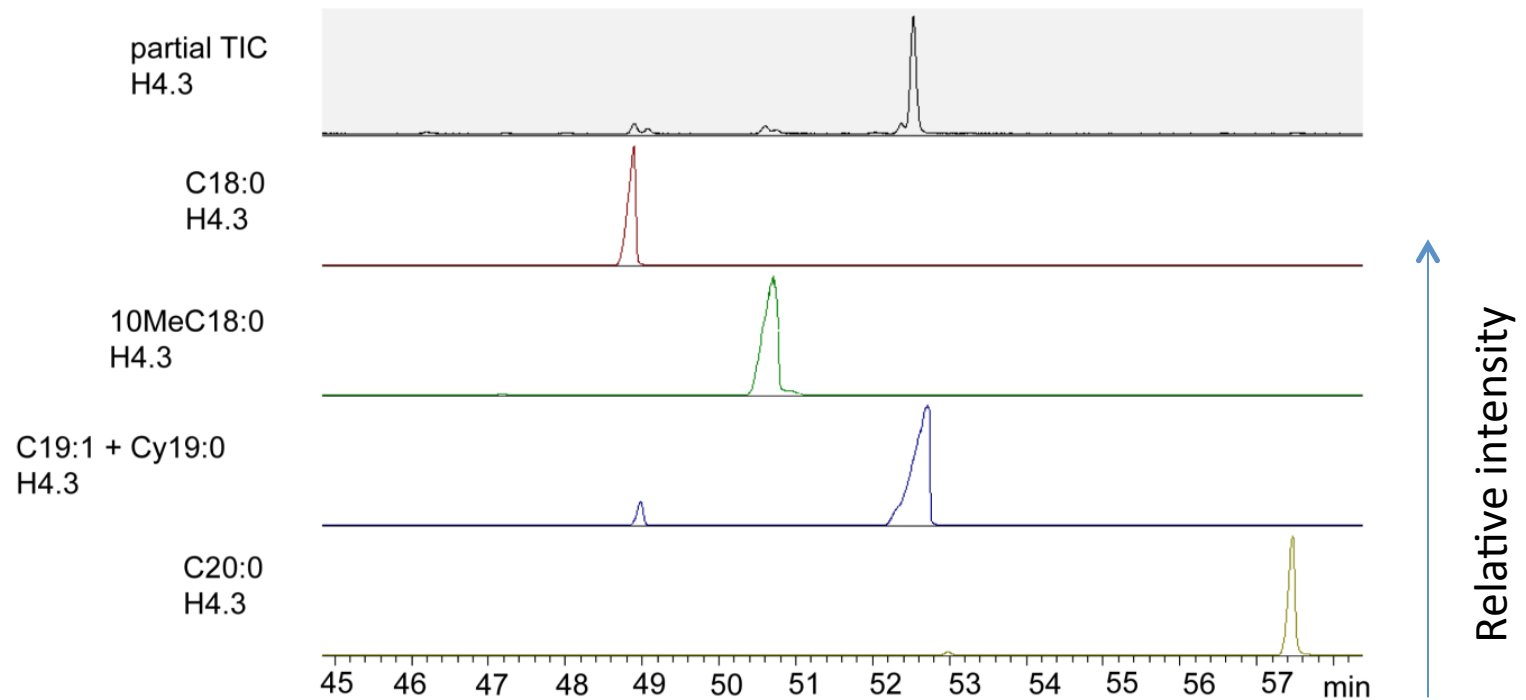
% ACN in
MeOH/H₂O
1/1



TIC (GC-MS) of the different collected HPLC fractions

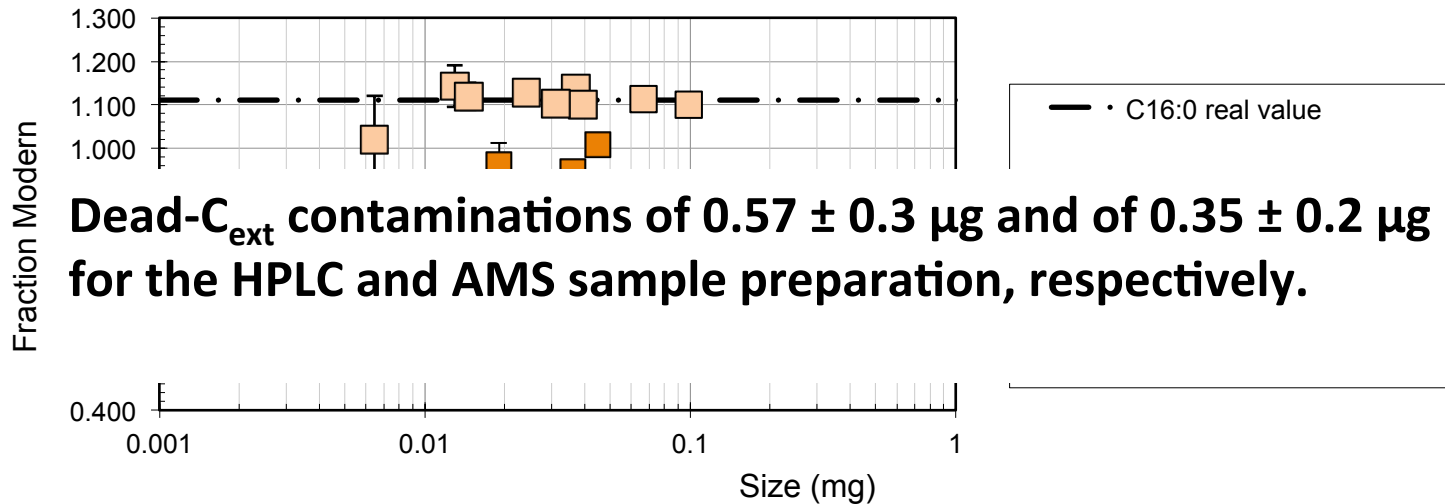


TIC (GC-MS) of the different collected HPLC fractions



HPLC column contamination

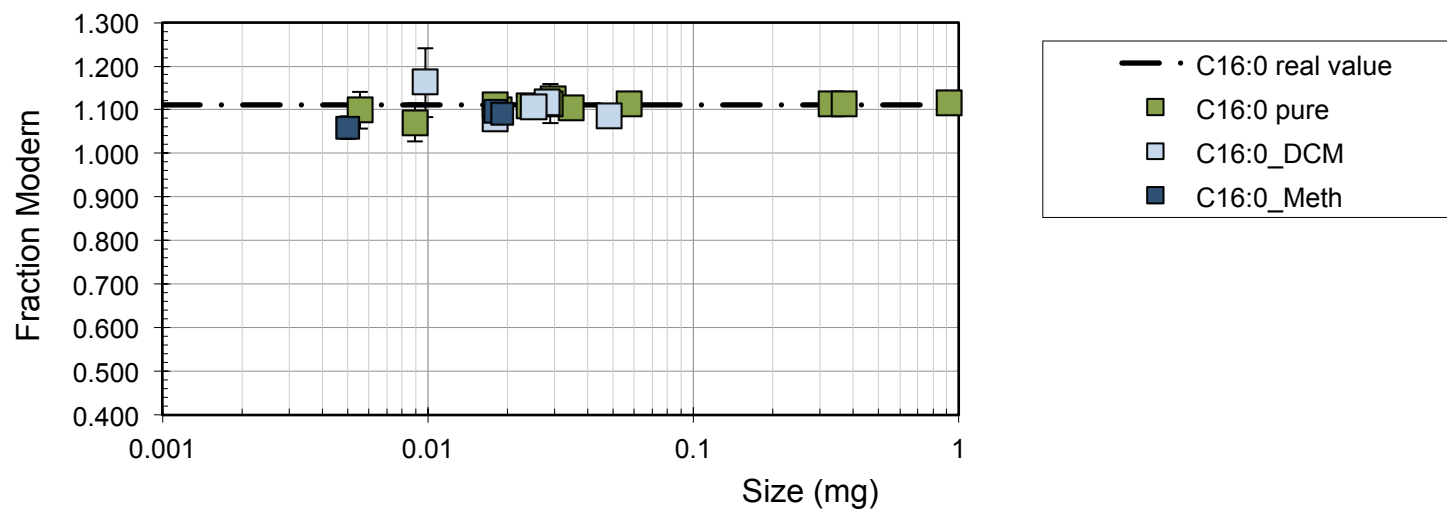
C16:0 FAME standard: $-132 \pm 3\text{‰}$



Phenanthrene standard: $-997 \pm 1\text{‰}$

Modern C_{ext} contamination was $0.40 \pm 0.20 \mu\text{g}$ for AMS sample preparation only.

Solvent contamination



- Each system will be distinct in term of Cext, therefore each user needs to evaluate the Cex and Fmex values specific for their system
- When considering CSRA applications, one must consider the magnitude of uncertainty required to provide useful information about the system being studied.

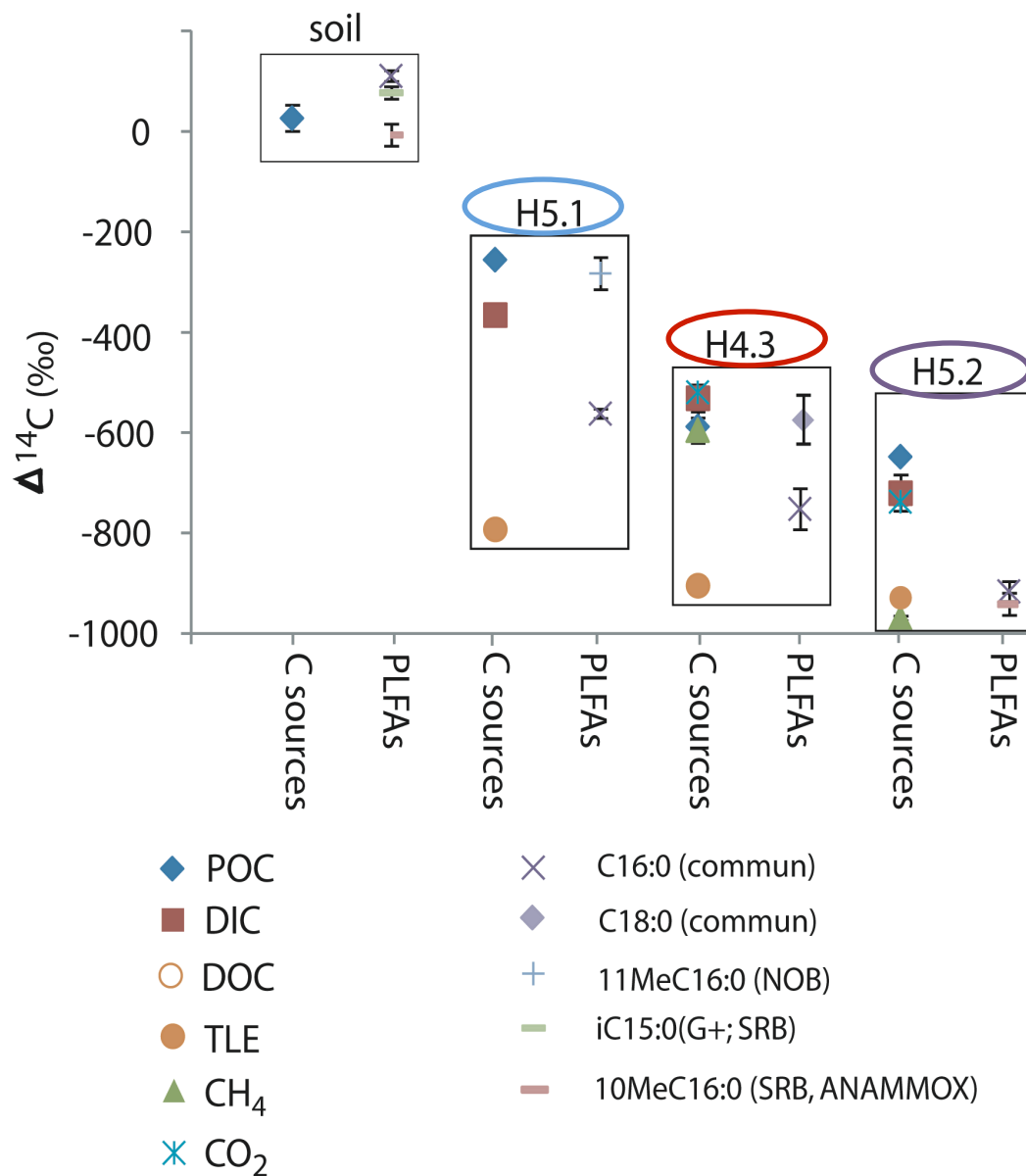
Mean recoveries

Table 2: Compound recovery in μC measured after the different purification and extraction steps

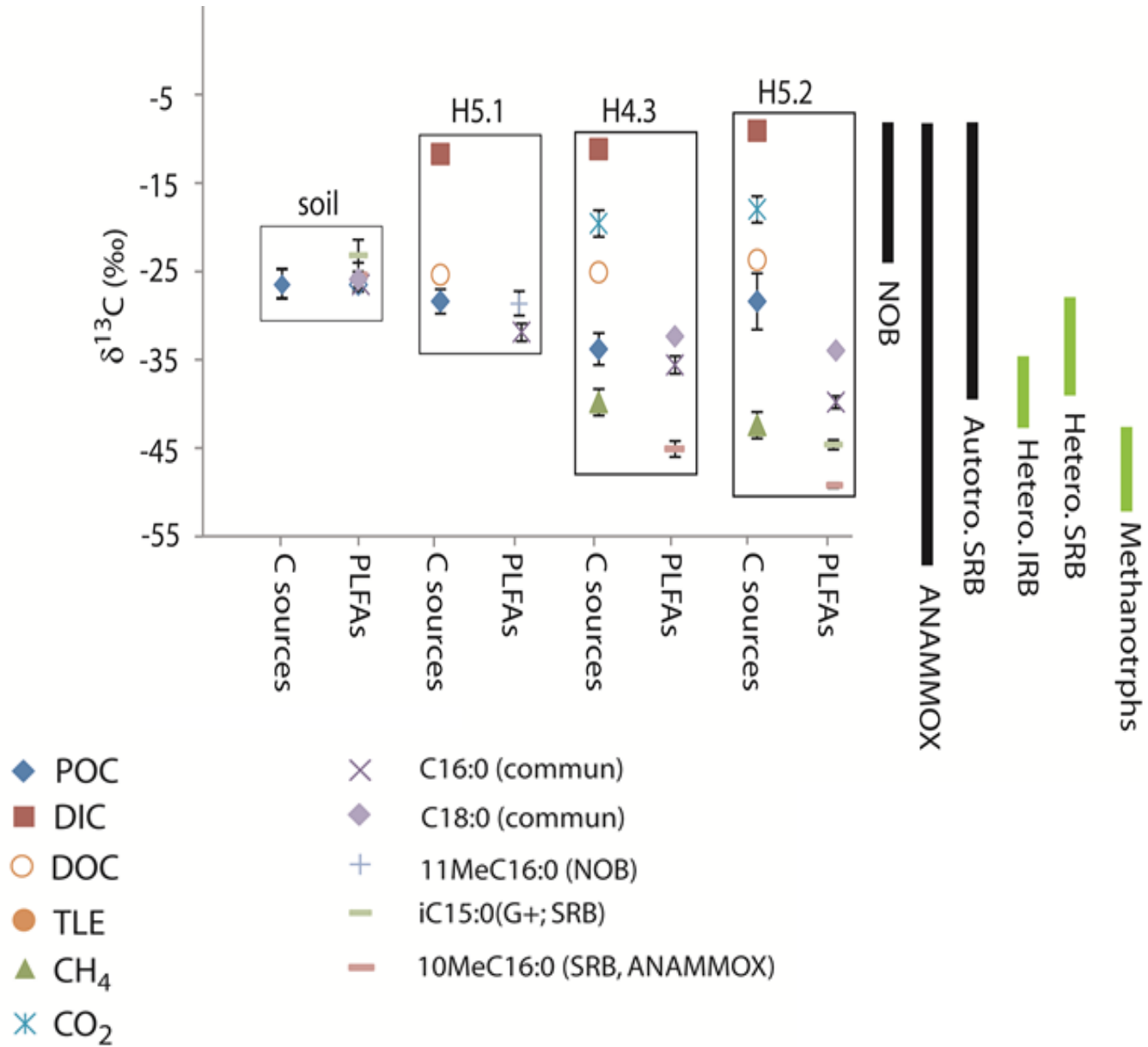
% recovery extraction + methylation + NH_2 column	% recovery of Ag-impregnated SCX column	% recovery of HPLC purification	% recovery of solvent evaporation and combustion	Total recovery
89 \pm 10 (2)	88 \pm 4 (9)	83 \pm 6 (9)	71 \pm 13 (9)	46 \pm 16 (9)

Larger compound lost during solvent evaporation or/and in vacuum line

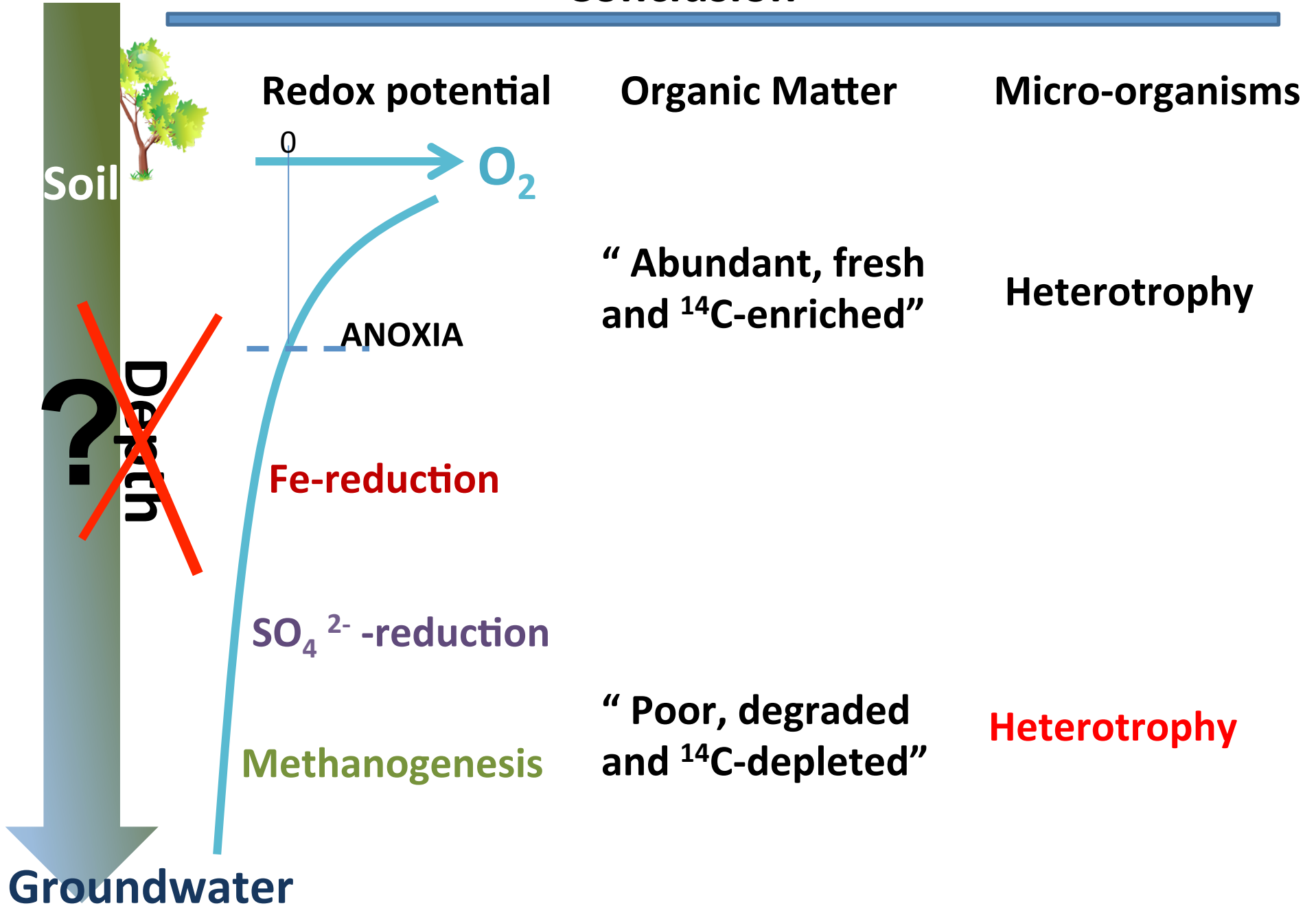
Results and implications



Results and implications



Conclusion





Acknowledgements

Susan Trumbore

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Georg Pohner

Jan Muhr

Kirsten Küsel

Kai U. Totsche

Vanessa-Nina Roth

Robert Lehmann

Heiko Minkmar

Nico Ueberschaar

...

Thank you for your attention

DFG Deutsche
Forschungsgemeinschaft



seit 1558

